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(54) Title: HUMAN TRANSCRIPTION FACTORS AND BINDING ASSAYS (57) Abstract The invention provides methods and compositions for identifying pharmacological agents useful in the diagnosis or treatment of disease associated with the expression of a gene modulated by a transcription complex containing at least a human nuclear factor of activated T-cells (hNFAT). The materials include a family of hNFAT proteins, active fragments thereof, and nucleic acids encoding them. The methods are particularly suited to high-throughput screening where one or more steps are performed by a computer controlled electromechanical robot comprising an axial rotatable arm.		

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Human Transcription Factors and Binding Assays

INTRODUCTION

Field of the Invention

The field of this invention is human transcription factors of activated T-cells.

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Background

Identifying and developing new pharmaceuticals is a multibillion dollar industry in the U.S. alone. Gene specific transcription factors provide a promising class of targets for novel therapeutics directed to these and other human diseases.

- 10 Urgently needed are efficient methods of identifying pharmacological agents or drugs which are active at the level of gene transcription. If amenable to automated, cost-effective, high throughput drug screening, such methods would have immediate application in a broad range of domestic and international pharmaceutical and biotechnology drug development programs.

- 15 Immunosuppression is therapeutically desirable in a wide variety of circumstances including transplantation, allergy and other forms of hypersensitivity, autoimmunity, etc. Cyclosporin, a widely used drug for effecting immunosuppression, is believed to act by inhibiting a calcineurin, a phosphatase which activates certain nuclear factors of activated T-cells (NFATs). However,
20 because of side effects and toxicity, clinical indications of cyclosporin (and the more recently developed FK506) are limited.

- Accordingly, it is desired to identify agents which more specifically interfere with the function of hNFATs. Unfortunately, the reagents necessary for the development of high-throughput screening assays for such therapeutics are
25 unavailable.

Relevant Literature

Nolan (June 17, 1994) Cell 77, 1-20 provides a recent review and commentary on molecular interactions of hNFAT proteins. Northrop et al. (June 9, 1994) Nature 369, 497-502 report the cloning of a cDNA encoding human NFATc. McCaffrey et al. (October 29, 1993) Science 262, 750-754 report the cloning of a fragment of a gene encoding a murine NFATp₁.

SUMMARY OF THE INVENTION

The invention provides methods and compositions for identifying lead compounds and pharmacological agents useful in the diagnosis or treatment of disease associated with the expression of one or more genes modulated by a transcription complex containing a human nuclear factor of activated T-cells (hNFAT). Several forms of hNFAT are provided including hNFATs designated hNFATp₁, hNFATp₂, hNFATc, hNFAT3, hNFAT4a, hNFAT4b and hNFAT4c. The invention also provides isolated nucleic acid encoding the subject hNFATs, vectors and cells comprising such nucleic acids, and methods of recombinantly producing polypeptides comprising hNFAT. The invention also provides hNFAT-specific binding reagents such as hNFAT-specific antibodies.

Methods using the disclosed hNFATs in drug development programs involve combining a selected hNFAT with a natural intracellular hNFAT binding target and a candidate pharmacological agent. Natural intracellular binding targets include transcription factors, such as AP1 proteins and nucleic acids encoding a hNFAT binding sequence. The resultant mixture is incubated under conditions whereby, but for the presence of the candidate pharmacological agent, the hNFAT selectively binds the target. Then the presence or absence of selective binding between the hNFAT and target is detected. A wide variety of alternative embodiments of the general methods using hNFATs are disclosed. The methods are particularly suited to high-throughput screening where one or more steps are performed by a computer controlled electromechanical robot comprising an axial rotatable arm and the solid substrate is a portion of a well of a microtiter plate.

hNFAT SEQUENCE ID NOS:

hNFATp ₁	cDNA	SEQUENCE ID NO:1
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	hNFATp ₁	protein	SEQUENCE ID NO:2
	hNFATp ₂	cDNA	SEQUENCE ID NO:1, bases 1-356 and 868-3478
	hNFATp ₂	protein	SEQUENCE ID NO:2, residues 220-1021
	hNFATc	cDNA	SEQUENCE ID NO:3
5	hNFATc	protein	SEQUENCE ID NO:4
	hNFAT3	cDNA	SEQUENCE ID NO:5
	hNFAT3	protein	SEQUENCE ID NO:6
	hNFAT4a	cDNA	SEQUENCE ID NO:7
	hNFAT4a	protein	SEQUENCE ID NO:8
10	hNFAT4b	cDNA	SEQUENCE ID NO:7, bases 211-2307 and SEQUENCE ID NO:9
	hNFAT4b	protein	SEQUENCE ID NO:8, residues 1-699 and SEQUENCE ID NO:10
	hNFAT4c	cDNA	SEQUENCE ID NO:7, bases 211-2307 and SEQUENCE ID NO:11
15	hNFAT4c	protein	SEQUENCE ID NO:8, residues 1-699 and SEQUENCE ID NO:12

DETAILED DESCRIPTION OF THE INVENTION

20 The invention provides methods and compositions relating to human NFATs. The subject hNFATs include regulators of cytokine gene expression that modulate immune system function. As such, hNFATs and hNFAT-encoding nucleic acids provide important targets for therapeutic intervention.

hNFATs derive from human cells, comprise invariant hNFAT rel domain
25 peptides (see, Table 1) and share at least 50% pair-wise rel sequence identity with each of the disclosed hNFAT sequences. Invariant hNFAT rel domain peptides include from the N-terminal end of the rel domain, HHRAHYETEGSRGAVKA (SEQUENCE ID NO:2, residues 419-435), PHAFYQVHRITGK (SEQUENCE ID NO:2, residues 470-482), IDCAGILKLRN (SEQUENCE ID NO:2, residues 513-
30 523), DIELRKGETDIGRKNTRVRLVFRVHX₁P (SEQUENCE ID NO:13), and PX₂ECSQRSAX₃ELP (SEQUENCE ID NO:14), where each X₁ and X₂ is hydrophobic residue such as valine or isoleucine, and X₃ is any residue, but preferably glutamine or histidine.

Table 1. hNFAT rel domains

NFATp (SEQ ID NO:2, residues 388-678)			
NFATc (SEQ ID NO:4, residues 406-697)			
5	NFAT3 (SEQ ID NO:6, residues 397-686)		
NFAT4b/c (SEQ ID NO:8, residues 411-702 and SEQ ID NO:10; SEQ ID NO:8, residues 411-702 and SEQ ID NO:12)			
10	NFATp	IPVTASLPPLDWPLSSQSGSYELRIEVQPKPHHRAHYETEGSRGAVKAPT	50
	NFATc	SYMSPTLPALDWQLPSHSGPYELRIEVQPKSHHRAHYETEGSRGAVKASA	50
	NFAT3	IFRTSALPPLDWPLPSQYEQLRLRIEVQPKRAHRAHYETEGSRGAVKAAP	50
	NFAT4b/c	IFRTSSLPPLDWPLPAHFGQCELKIEVQPKTHHRAHYETEGSRGAVKAST	50
15	NFATp	GGHPVVQLHGYMENKPLGLQIFIGTADERILKPHAFYQVHRITGKTVTTT	100
	NFATc	GGHPVQLHGYLENEPLMLQLFIGTADDRLLRPHAFYQVHRITGKTVSTT	100
	NFAT3	GGHPVVKLLGYS-EKPLTLQMPIGTADERNLRPHAFYQVHRITGKTIVATA	99
	NFAT4b/c	GGHPVVKLLGYN-EKPINLQMPIGTADDRYL RPHAFYQVHRITGKTIVATA	99
20	NFATp	SYEKIVGNTKVLLEIPLEPKNNMRATIDCAGILKLRNADIELRKGETDIGR	150
	NFATc	SHEAILSNTKVLLEIPLLPENSMRAVIDCAGILKLRNSDIELRKGETDIGR	150
	NFAT3	SYEAVVSGTKVLEMTLLPENMAANIDCAGILKLRNSDIELRKGETDIGR	149
	NFAT4b/c	SQEIIIASTKVLLEIPLLPENMSASIDCAGILKLRNSDIELRKGETDIGR	149
25	NFATp	KNTRVRLVFRVHIEPSSGRIVSLQTASNPIECSQRSAHELPMVERQDSTD	200
	NFATc	KNTRVRLVFRVHVPQPSGRTLSLQVASNPIECSQRSAQELPLVEKQSTD	200
	NFAT3	KNTRVRLVFRVHVPQGGKVVSVQAASVPIECSQRSAQELPQVEAYSPSA	199
	NFAT4b/c	KNTRVRLVFRVHVPQPSGKVLSLQIASIPVECSQRSAQELPHIEKYSINS	199
30	NFATp	CLVYGGQQMILTQGNFTSESKVVFTEKTTDQQQIWEMEATVDKDKSQPNM	250
	NFATc	YPVVGGKKMVLSCGNFLQDSKVIFVEKAPDGHVWEMEAKTDRDLCKPNS	250
	NFAT3	CSVRCGEELVLTGSHFLPDSKVVFIERGPDGKLQWEEATVNRLQSNEVT	249
	NFAT4b/c	CSVNGGHEMVVTGSHFLPESKIIIFLEKGQDGRPQWEVEGKIIREKCCGAH	249
35	NFATp	LFVEIPEYRNKHIRTPVKVNFYVINGKRRRSQPQHFTYHPV	291
	NFATc	LVVEIPEFRNQRITSPVHVSFYVCNGKRRRSQYQRFYTLPA	291
	NFAT3	LTLTVPEYSMKRVSRPVQVYFYVSNGRRRSPTQSFRFLPV	290
	NFAT4b/c	IVLEVPPYHNPVATAAVOVHFYLCNGKRRRSQSQRFTYTFV	290

In addition to the shared rel domains, some hNFATs have smaller regions of
 40 sequence similarity on the terminal side of the rel domains. For example, the amino
 terminal regions of hNFAT 4a, 4b and 4c and hNFATc have several regions of
 similarity (Table 2). The two largest regions (designated regions A and B in Table 2)
 contain 23 of 41 and 24 of 45 identical amino acids between the two proteins.
 hNFATp and hNFAT3 also have similarity to other hNFAT proteins in this region
 45 (Table 2). The homology between hNFAT3 and hNFAT 4a, 4b and 4c extends about
 25 amino acids upstream of the rel region (designated region C in Table 2).

Table 2. hNFAT regions 5' to the rel domain

50	A	NFATc	PSTATLSLPSLEAYRDPB-CLSPASSLSRRSCNSEASSYES	195
		NFAT4	PBRDHLYLPLEPSYRESSLSPPASSISSRSWFSDASSCES	189

		NFATc (SEQ ID NO:4, residues 152-191)	
		NFAT4a (SEQ ID NO:8, residues 144-184)	
5	[NFATc SPQHSPSTSPRASVTEESWLGAR-----SSRPASPCNKRKYSLNG	272
		NFAT4 SPRQSPCHSPRSSVTDENWLSPPASGPPSRPTSPCGKRRSSAEV	281
		NFATc (SEQ ID NO:4, residues 233-272)	
		NFAT4a (SEQ ID NO:8, residues 236-281)	
B	[NFATc SSRPASPCNKRKYSLNG	272
		NFAT3 SPRPASPCGKRRYSSSG	275
10		NFATc (SEQ ID NO:4, residues 256-272)	
		NFAT3 (SEQ ID NO:6, residues 259-275)	
	[NFATc SPQHSPSTSPRASVTEESWLGARSSRP	272
		NFATp SPRTSPIMSPRTSLAEDSCLGRHSPVP	239
	L	NFATc (SEQ ID NO:4, residues 233-259)	
15		NFATp (SEQ ID NO:2, residues 213-239)	
C	[NFAT3 RKEVAGMDYLAVPSPPLAWSKARIQGHSP	396
		NFAT4 KKDSCGDQFLSVSPFTWSKPKPG-HTP	410
20	L	NFAT3 (SEQ ID NO:6, residues 369-396)	
		NFAT4a (SEQ ID NO:8, residues 384-410)	

Nucleic acids encoding hNFATs may be isolated from human cells by screening cDNA libraries for human immune cells with probes or PCR primers derived from the disclosed hNFAT genes. In addition to the invariant hNFAT rel sequences and the 50% pair-wise rel domain identity, cDNAs of hNFAT transcripts typically share substantially overall sequence identity with one or more of the disclosed hNFAT sequences.

The subject hNFAT fragments have one or more hNFAT-specific binding affinities, including the ability to specifically bind at least one natural human intracellular hNFAT-specific binding target or a hNFAT-specific binding agent such as a hNFAT-specific antibody or a hNFAT-specific binding agent identified in assays such as described below. Accordingly, the specificity of hNFAT fragment specific binding agents is confirmed by ensuring non-cross-reactivity with other NFATs. Furthermore, preferred hNFAT fragments are capable of eliciting an antibody capable of specifically binding an hNFAT. Methods for making immunogenic peptides through the use of conjugates, adjuvants, etc. and methods for eliciting antibodies, e.g. immunizing rabbits, are well known.

Exemplary natural intracellular binding targets include nucleic acids which comprise one or more hNFAT DNA binding sites. Functional hNFAT binding sites have been found in the promoters or enhancers of several different cytokine genes including IL-2, IL-4, IL-3, GM-CSF, and TNF- α and are often located next to AP-1

binding sites, which are recognized by members of the fos and jun families of transcription factors. Typically, the AP-1 binding sites adjacent to hNFAT sites are low affinity sites, and AP-1 proteins cannot bind them independently. However, many NF-AT and AP-1 protein combinations are capable of cooperatively binding to
5 DNA. Furthermore, cell-type specificity of cytokine gene transcription is often controlled, at least in part, by the combinations of hNFAT and AP-1 proteins present in those cells. For example, there are different classes of T cells that secrete different sets of cytokines: e.g. TH1 cells produce IL-2 and IFN- γ , while TH2 cells produce IL-4, IL-5, and IL-6. hNFAT binding sites are involved in the regulation of both TH1 and
10 TH2 cytokines. Further, differential expression of the cytokine gene in T cell subsets is controlled the combinatorial interactions of hNFAT and AP-1 proteins.

In addition to DNA binding sites and other transcription factors such as AP1, other natural intracellular binding targets include cytoplasmic proteins such as ankyrin repeat containing hNFAT inhibitors, protein serine/threonine kinases, etc., and
15 fragments of such targets which are capable of hNFAT-specific binding. Other natural hNFAT binding targets are readily identified by screening cells, membranes and cellular extracts and fractions with the disclosed materials and methods and by other methods known in the art. For example, two-hybrid screening using hNFAT fragments are used to identify intracellular targets which specifically bind such
20 fragments. Preferred hNFAT fragments retain the ability to specifically bind at least one of an hNFAT DNA binding site and can preferably cooperatively bind with AP1. Convenient ways to verify the ability of a given hNFAT fragment to specifically bind such targets include in vitro labelled binding assays such as described below, and EMSAs.

25 A wide variety of molecular and biochemical methods are available for generating and expressing hNFAT fragments, see e.g. Molecular Cloning, A Laboratory Manual (2nd Ed., Sambrook, Fritsch and Maniatis, Cold Spring Harbor), Current Protocols in Molecular Biology (Eds. Ausubel, Brent, Kingston, More, Feidman, Smith and Stuhl, Greene Publ. Assoc., Wiley-Interscience, NY, NY, 1992)
30 or that are otherwise known in the art. For example, hNFAT or fragments thereof may be obtained by chemical synthesis, expression in bacteria such as E. coli and eukaryotes such as yeast or vaccinia or baculovirus-based expression systems, etc., depending on the size, nature and quantity of the hNFAT or fragment. The subject

hNFAT fragments are of length sufficient to provide a novel peptide. As used herein, such peptides are at least 5, usually at least about 6, more usually at least about 8, most usually at least about 10 amino acids. hNFAT fragments may be present in a free state or bound to other components such as blocking groups to chemically
5 insulate reactive groups (e.g. amines, carboxyls, etc.) of the peptide, fusion peptides or polypeptides (i.e. the peptide may be present as a portion of a larger polypeptide), etc.

The subject hNFAT fragments maintain binding affinity of not less than six, preferably not less than four, more preferably not less than two orders of magnitude less than the binding equilibrium constant of a full-length native hNFAT to the
10 binding target under similar conditions. Particular hNFAT fragments or deletion mutants are shown to function in a dominant-negative fashion. Such fragments provide therapeutic agents, e.g. when delivered by intracellular immunization - transfection of susceptible cells with nucleic acids encoding such mutants.

The claimed hNFAT and hNFAT fragments are isolated, partially pure or pure
15 and are typically recombinantly produced. As used herein, an "isolated" peptide is unaccompanied by at least some of the material with which it is associated in its natural state and constitutes at least about 0.5%, preferably at least about 2%, and more preferably at least about 5% by weight of the total protein (including peptide) in a given sample; a partially pure peptide constitutes at least about 10% , preferably at
20 least about 30%, and more preferably at least about 60% by weight of the total protein in a given sample; and a pure peptide constitutes at least about 70% , preferably at least about 90%, and more preferably at least about 95% by weight of the total protein in a given sample.

Preferred hNFAT fragments comprise at least a functional portion of the rel
25 domain. There are several different biochemical functions that are mediated by the rel and hNFAT rel-similarity domains: DNA binding, dimerization, interaction with B-zip proteins, interaction with inhibitor proteins, and nuclear localization. Other rel family proteins have been shown to physically interact with AP-1 (fos and jun) proteins (Stein et al., EMBO J. 12, 1993). The rel homology domain is necessary for
30 this interaction and the B-zip region of the AP-1 proteins is involved in this protein-protein interaction. The specificity in the ability of hNFAT and AP-1 family members to interact is related to the tissue specific and cell type specific regulation of gene expression governed by these proteins. The rel and rel-similarity domains also

interact with members of the I- κ B family of inhibitor proteins including I- κ B-like ankyrin repeat proteins (reviewed in Beg and Baldwin, Genes and Dev., 1993). The C-terminal half or the rel domain is involved the interaction with I- κ B. There are 5 related I- κ B-like proteins which are characterized by having multiple copies of a 33 amino acid sequence motif called the ankyrin repeat.

The invention provides hNFAT-specific binding agents, methods of identifying and making such agents, and their use in diagnosis, therapy and pharmaceutical development. For example, hNFAT-specific agents are useful in a variety of diagnostic applications, especially where disease or disease prognosis is associated with immune dysfunction resulting from improper expression of hNFAT. Novel hNFAT-specific binding agents include hNFAT-specific antibodies and other natural intracellular binding agents identified with assays such as one- and two-hybrid screens; non-natural intracellular binding agents identified in screens of chemical libraries, etc.

Generally, hNFAT-specificity of the binding target is shown by binding equilibrium constants. Such targets are capable of selectively binding a hNFAT, i.e. with an equilibrium constant at least about 10^4 M^{-1} , preferably at least about 10^6 M^{-1} , more preferably at least about 10^8 M^{-1} . A wide variety of cell-based and cell-free assays may be used to demonstrate hNFAT-specific binding. Cell based assays include one and two-hybrid screens, mediating or competitively inhibiting hNFAT-mediated transcription, etc. Preferred are rapid in vitro, cell-free assays such as mediating or inhibiting hNFAT-protein (e.g. hNFAT-AP1 binding), hNFAT-nucleic acid binding, immunoassays, etc. Other useful screening assays for hNFAT/hNFAT fragment-target binding include fluorescence resonance energy transfer (FRET), electrophoretic mobility shift analysis (EMSA), etc.

The invention also provides nucleic acids encoding the subject hNFAT and hNFAT fragments, which nucleic acids may be part of hNFAT-expression vectors and may be incorporated into recombinant cells for expression and screening, transgenic animals for functional studies (e.g. the efficacy of candidate drugs for disease associated with expression of a hNFAT), etc. In addition, the invention provides nucleic acids sharing substantial sequence similarity with that of one or more wild-type hNFAT nucleic acids. Substantially identical or homologous nucleic acid

sequences hybridize to their respective complements under high stringency conditions, for example, at 55°C and hybridization buffer comprising 50% formamide in 0.9 M saline/0.09 M sodium citrate (SSC) buffer and remain bound when subject to washing at 55°C with the SSC/formamide buffer. Where the sequences diverge, the
5 differences are preferably silent, i.e. or a nucleotide change providing a redundant codon, or conservative, i.e. a nucleotide change providing a conservative amino acid substitution.

The subject nucleic acids find a wide variety of applications including use as hybridization probes, PCR primers, therapeutic nucleic acids, etc. for use in detecting
10 the presence of hNFAT genes and gene transcripts, for detecting or amplifying nucleic acids with substantial sequence similarity such as hNFAT homologs and structural analogs, and for gene therapy applications. Given the subject probes, materials and methods for probing cDNA and genetic libraries and recovering homologs are known in the art. Preferred libraries are derived from human immune cells, especially cDNA
15 libraries from differentiated and activated human lymphoid cells. In one application, the subject nucleic acids find use as hybridization probes for identifying hNFAT cDNA homologs with substantial sequence similarity. These homologs in turn provide additional hNFATs and hNFAT fragments for use in binding assays and therapy as described herein. hNFAT encoding nucleic acids also find applications in
20 gene therapy. For example, nucleic acids encoding dominant-negative hNFAT mutants are cloned into a virus and the virus used to transfect and confer disease resistance to the transfected cells..

Therapeutic hNFAT nucleic acids are used to modulate, usually reduce, cellular expression or intracellular concentration or availability of active hNFAT.
25 These nucleic acids are typically antisense: single-stranded sequences comprising complements of the disclosed hNFAT nucleic acids. Antisense modulation of hNFAT expression may employ hNFAT antisense nucleic acids operably linked to gene regulatory sequences. Cells are transfected with a vector comprising an hNFAT sequence with a promoter sequence oriented such that transcription of the gene yields
30 an antisense transcript capable of binding to endogenous hNFAT encoding mRNA. Transcription of the antisense nucleic acid may be constitutive or inducible and the vector may provide for stable extrachromosomal maintenance or integration. Alternatively, single-stranded antisense nucleic acids that bind to genomic DNA or

mRNA encoding a hNFAT or hNFAT fragment may be administered to the target cell, in or temporarily isolated from a host, at a concentration that results in a substantial reduction in hNFAT expression. For gene therapy involving the transfusion of hNFAT transfected cells, administration will depend on a number of variables that are ascertained empirically. For example, the number of cells will vary depending on the stability of the transfused cells. Transfusion media is typically a buffered saline solution or other pharmacologically acceptable solution. Similarly the amount of other administered compositions, e.g. transfected nucleic acid, protein, etc., will depend on the manner of administration, purpose of the therapy, and the like.

10 The subject nucleic acids are often recombinant, meaning they comprise a sequence joined to a nucleotide other than that which it is joined to on a natural chromosome. An isolated nucleic acid constitutes at least about 0.5% , preferably at least about 2%, and more preferably at least about 5% by weight of total nucleic acid present in a given fraction. A partially pure nucleic acid constitutes at least about
15 10%, preferably at least about 30%, and more preferably at least about 60% by weight of total nucleic acid present in a given fraction. A pure nucleic acid constitutes at least about 80%, preferably at least about 90%, and more preferably at least about 95% by weight of total nucleic acid present in a given fraction.

 The invention provides efficient methods of identifying pharmacological
20 agents or drugs which are active at the level of hNFAT modulatable cellular function, particularly hNFAT mediated interleukin signal transduction. Generally, these screening methods involve assaying for compounds which interfere with hNFAT activity such as hNFAT-AP1 binding, hNFAT-DNA binding, etc. The methods are amenable to automated, cost-effective high throughput drug screening and have
25 immediate application in a broad range of domestic and international pharmaceutical and biotechnology drug development programs.

 Target therapeutic indications are limited only in that the target cellular function (e.g. gene expression) be subject to modulation, usually inhibition, by disruption of the formation of a complex (e.g. transcription complex) comprising a
30 hNFAT or hNFAT fragment and one or more natural hNFAT intracellular binding targets. Since a wide variety of genes are subject to hNFAT regulated gene transcription, target indications may include infection, metabolic disease, genetic disease, cell growth and regulatory disfunction, such as neoplasia, inflammation.

hypersensitivity, etc. Frequently, the target indication is related to either immune dysfunction or selective immune suppression.

A wide variety of assays for binding agents are provided including labelled in vitro protein-protein and protein-DNA binding assay, electrophoretic mobility shift
5 assays, immunoassays for protein binding or transcription complex formation, cell based assays such as one, two and three hybrid screens, expression assays such as transcription assays, etc. For example, three-hybrid screens are used to rapidly examine the effect of transfected nucleic acids, which may, for example, encode combinatorial peptide libraries or antisense molecules, on the intracellular binding of
10 hNFAT or hNFAT fragments to intracellular hNFAT targets. Convenient reagents for such assays (e.g. GAL4 fusion partners) are known in the art.

hNFAT or hNFAT fragments used in the methods are usually added in an isolated, partially pure or pure form and are typically recombinantly produced. The hNFAT or fragment may be part of a fusion product with another peptide or
15 polypeptide, e.g. a polypeptide that is capable of providing or enhancing protein-protein binding, sequence-specific nucleic acid binding or stability under assay conditions (e.g. a tag for detection or anchoring).

The assay mixtures comprise at least a portion of a natural intracellular hNFAT binding target such as AP1 or a nucleic acid comprising a sequence which
20 shares sufficient sequence similarity with a gene or gene regulatory region to which the native hNFAT naturally binds to provide sequence-specific binding of the hNFAT or hNFAT fragment. Such a nucleic acid may further comprise one or more sequences which facilitate the binding of a second transcription factor or fragment thereof which cooperatively binds the nucleic acid with the hNFAT (i.e. at least one
25 increases the affinity or specificity of the DNA binding of the other). While native binding targets may be used, it is frequently preferred to use portions (e.g. peptides, nucleic acid fragments) or analogs (i.e. agents which mimic the hNFAT binding properties of the natural binding target for the purposes of the assay) thereof so long as the portion provides binding affinity and avidity to the hNFAT conveniently
30 measurable in the assay. Binding sequences for other transcription factors may be found in sources such as the Transcription Factor Database of the National Center for Biotechnology Information at the National Library for Medicine, in Faisst and Meyer (1991) Nucleic Acids Research 20, 3-26, and others known to those skilled in this art.

Where used, the nucleic acid portion bound by the peptide(s) may be continuous or segmented and is usually linear and double-stranded DNA, though circular plasmids or other nucleic acids or structural analogs may be substituted so long as hNFAT sequence-specific binding is retained. In some applications, supercoiled DNA provides optimal sequence-specific binding and is preferred. The nucleic acid may be of any length amenable to the assay conditions and requirements. Typically the nucleic acid is between 8 bp and 5 kb, preferably between about 12 bp and 1 kb, more preferably between about 18 bp and 250 bp, most preferably between about 27 and 50 bp. Additional nucleotides may be used to provide structure which enhances or decreased binding or stability, etc. For example, combinatorial DNA binding can be effected by including two or more DNA binding sites for different or the same transcription factor on the oligonucleotide. This allows for the study of cooperative or synergistic DNA binding of two or more factors. In addition, the nucleic acid can comprise a cassette into which transcription factor binding sites are conveniently spliced for use in the subject assays.

The assay mixture also comprises a candidate pharmacological agent. Generally a plurality of assay mixtures are run in parallel with different agent concentrations to obtain a differential response to the various concentrations. Typically, one of these concentrations serves as a negative control, i.e. at zero concentration or below the limits of assay detection. Candidate agents encompass numerous chemical classes, though typically they are organic compounds; preferably small organic compounds. Small organic compounds have a molecular weight of more than 50 yet less than about 2,500, preferably less than about 1000, more preferably, less than about 500. Candidate agents comprise functional chemical groups necessary for structural interactions with proteins and/or DNA, and typically include at least an amine, carbonyl, hydroxyl or carboxyl group, preferably at least two of the functional chemical groups, more preferably at least three. The candidate agents often comprise cyclical carbon or heterocyclic structures and/or aromatic or polyaromatic structures substituted with one or more of the forementioned functional groups. Candidate agents are also found among biomolecules including peptides, saccharides, fatty acids, steroids, purines, pyrimidines, derivatives, structural analogs or combinations thereof, and the like. Where the agent is or is encoded by a transfected nucleic acid, said nucleic acid is typically DNA or RNA.

Candidate agents are obtained from a wide variety of sources including libraries of synthetic or natural compounds. For example, numerous means are available for random and directed synthesis of a wide variety of organic compounds and biomolecules, including expression of randomized oligonucleotides.

5 Alternatively, libraries of natural compounds in the form of bacterial, fungal, plant and animal extracts are available or readily produced. Additionally, natural and synthetically produced libraries and compounds are readily modified through conventional chemical, physical, and biochemical means. In addition, known pharmacological agents may be subject to directed or random chemical modifications,

10 such as acylation, alkylation, esterification, amidification, etc., to produce structural analogs.

A variety of other reagents may also be included in the mixture. These include reagents like salts, buffers, neutral proteins, e.g. albumin, detergents, etc. which may be used to facilitate optimal protein-protein and/or protein-nucleic acid binding and/or

15 reduce non-specific or background interactions, etc. Also, reagents that otherwise improve the efficiency of the assay, such as protease inhibitors, nuclease inhibitors, antimicrobial agents, etc. may be used.

The resultant mixture is incubated under conditions whereby, but for the presence of the candidate pharmacological agent, the hNFAT specifically binds the

20 cellular binding target, portion or analog. The mixture components can be added in any order that provides for the requisite bindings. Incubations may be performed at any temperature which facilitates optimal binding, typically between 4 and 40 °C, more commonly between 15 and 40°C. Incubation periods are likewise selected for optimal binding but also minimized to facilitate rapid, high-throughput screening, and

25 are typically between .1 and 10 hours, preferably less than 5 hours, more preferably less than 2 hours.

After incubation, the presence or absence of specific binding between the hNFAT and one or more binding targets is detected by any convenient way. For cell-free binding type assays, a separation step is often used to separate bound from

30 unbound components. The separation step may be accomplished in a variety of ways. Conveniently, at least one of the components is immobilized on a solid substrate which may be any solid from which the unbound components may be conveniently separated. The solid substrate may be made of a wide variety of materials and in a

wide variety of shapes, e.g. microtiter plate, microbead, dipstick, resin particle, etc. The substrate is chosen to maximize signal to noise ratios, primarily to minimize background binding, for ease of washing and cost.

Separation may be effected for example, by removing a bead or dipstick from
5 a reservoir, emptying or diluting reservoir such as a microtiter plate well, rinsing a bead (e.g. beads with iron cores may be readily isolated and washed using magnets), particle, chromatographic column or filter with a wash solution or solvent. Typically, the separation step will include an extended rinse or wash or a plurality of rinses or washes. For example, where the solid substrate is a microtiter plate, the wells may be
10 washed several times with a washing solution, which typically includes those components of the incubation mixture that do not participate in specific binding such as salts, buffer, detergent, nonspecific protein, etc. may exploit a polypeptide specific binding reagent such as an antibody or receptor specific to a ligand of the polypeptide.

Detection may be effected in any convenient way. For cell based assays such
15 as one, two, and three hybrid screens, the transcript resulting from hNFAT-target binding usually encodes a directly or indirectly detectable product (e.g. galactosidase activity, luciferase activity, etc.). For cell-free binding assays, one of the components usually comprises or is coupled to a label. A wide variety of labels may be employed - essentially any label that provides for detection of bound protein. The label may
20 provide for direct detection as radioactivity, luminescence, optical or electron density, etc. or indirect detection such as an epitope tag, an enzyme, etc. The label may be appended to the protein e.g. a phosphate group comprising a radioactive isotope of phosphorous, or incorporated into the protein structure, e.g. a methionine residue comprising a radioactive isotope of sulfur.

25 A variety of methods may be used to detect the label depending on the nature of the label and other assay components. For example, the label may be detected bound to the solid substrate or a portion of the bound complex containing the label may be separated from the solid substrate, and thereafter the label detected. Labels may be directly detected through optical or electron density, radiative emissions,
30 nonradiative energy transfers, etc. or indirectly detected with antibody conjugates, etc. For example, in the case of radioactive labels, emissions may be detected directly, e.g. with particle counters or indirectly, e.g. with scintillation cocktails and counters. The methods are particularly suited to automated high throughput drug screening.

Candidate agents shown to inhibit hNFAT - target binding or transcription complex formation provide valuable reagents to the pharmaceutical industries for animal and human trials.

As previously described, the methods are particularly suited to automated high
5 throughput drug screening. In a particular embodiment, the arm retrieves and transfers a microtiter plate to a liquid dispensing station where measured aliquots of each an incubation buffer and a solution comprising one or more candidate agents are deposited into each designated well. The arm then retrieves and transfers to and deposits in designated wells a measured aliquot of a solution comprising a labeled
10 transcription factor protein. After a first incubation period, the liquid dispensing station deposits in each designated well a measured aliquot of a biotinylated nucleic acid solution. The first and/or following second incubation may optionally occur after the arm transfers the plate to a shaker station. After a second incubation period, the arm transfers the microtiter plate to a wash station where the unbound contents of each
15 well is aspirated and then the well repeatedly filled with a wash buffer and aspirated. Where the bound label is radioactive phosphorous, the arm retrieves and transfers the plate to the liquid dispensing station where a measured aliquot of a scintillation cocktail is deposited in each designated well. Thereafter, the amount of label retained in each designated well is quantified.

20 In more preferred embodiments, the liquid dispensing station and arm are capable of depositing aliquots in at least eight wells simultaneously and the wash station is capable of filling and aspirating ninety-six wells simultaneously. Preferred robots are capable of processing at least 640 and preferably at least about 1,280 candidate agents every 24 hours, e.g. in microtiter plates. Of course, useful agents are
25 identified with a range of other assays (e.g. gel shifts, etc.) employing the subject hNFAT and hNFAT fragments.

The subject hNFAT and hNFAT fragments and nucleic acids provide a wide variety of uses in addition to the in vitro binding assays described above. For example, cell-based assays are provided which involve transfecting a T-cell antigen
30 receptor expressing cell with an hNFAT inducible reporter such as luciferase. Agents which modulate hNFAT mediated cell function are then detected through a change in the reporter.

The following examples are offered by way of illustration and not by way of limitation.

EXPERIMENTAL

- Investigation of the antigen inducible expression of the IL-2 gene led to the discovery of the regulatory transcription factor NFAT (Nuclear Factor of Activated T cells) (Durand et al. 1988; Shaw et al. 1988). Like several other transcription factors involved in mediating signal transduction, the activity of NFAT is regulated by subcellular localization. In resting T cells NFAT activity is restricted to cytoplasm; stimulation of the T cell receptor leads to translocation of NFAT to the nucleus.
- 10 Movement of NFAT to the nucleus is dependent on the activation of the calcium-regulated phosphatase calcineurin (Clippstone and Crabtree 1992). The immunosuppressive drugs cyclosporin and FK506 inhibit the activity of calcineurin, and thereby prevent the nuclear localization of NFAT and subsequent activation of cytokine gene expression (reviewed in (Schreiber and Crabtree 1992).
- 15 Activation of the T cell antigen receptor induces two signalling pathways required for IL-2 induction, one is the cyclosporin-sensitive, calcium-dependent pathway and the other relies on the activation of protein kinase C (PKC). Antigenic stimulation of these pathways can be mimicked by treating cells with a calcium ionophore and a phorbol ester. The PKC-inducible activity was found to be mediated
- 20 by fos and jun proteins (Jain et al. 1992; Northrop et al. 1993). The NFAT binding site in the IL-2 promoter is adjacent to a weak binding site for AP-1 proteins, and NFAT and AP-1 proteins bind cooperatively to this composite element (Jain et al. 1993; Northrop et al. 1993). The transcriptional activation mediated by AP-1 proteins through this site appears to be critical for IL-2 expression in activated T cells.
- 25 There are several different combinations of fos and jun family members that can interact with NFAT to bind DNA (Boise et al. 1993; Northrop et al. 1993; Jain et al. 1994; Yaseen et al. 1994). Therefore, the composition of the AP-1 complex that interacts with NFAT may vary in different cell types and different stages of T cell activation. NFAT was originally reported to be a T cell specific transcription factor
- 30 critical for the restricted expression of IL-2 (Shaw et al. 1988). More recently, NFAT activity was detected in B cells (Brabletz et al. 1991; Yaseen et al. 1993; Choi et al. 1994; Venkataraman et al. 1994). This is consistent with the finding that, in

transgenic mice, the major sites of expression of a reporter gene regulated by the IL-2 NFAT/AP-1 site are activated T and B cells (Verweij et al. 1990).

In addition to IL-2, NFAT sites have been discovered in the promoters of several other cytokine genes, including IL-4 (Chuvpilo et al. 1993; Szabo et al. 1993; Rooney et al. 1994), IL-3 (Cockerill et al. 1993), GM-CSF (Masuda et al. 1993), and TNF- α (Goldfeld et al. 1993). Thus, it appears that NFAT proteins are involved in the coordinate regulation of many different cytokines in activated lymphocytes. As with IL-2, most of the NFAT sites in other cytokine promoters are composite elements that also contain AP-1 binding sites (Rao, 1994).

Distinct genes encoding NFAT proteins have now been isolated (Jain et al. 1993; McCaffrey et al. 1993; Northrop et al. 1994; Hoey et al., in press). Two of these genes, designated NFATp and NFATc, encode related proteins that are highly similar to each other within a 290 amino acid domain. This NFAT homology region shares weak sequence similarity with the DNA binding and dimerization domain of the rel family of transcription factors (reviewed in (Nolan 1994). There is evidence that both NFATp and NFATc may be involved in mediating transcriptional regulation in activated T cells. For example, NFATp forms a specific complex on DNA with fos and jun that activates transcription in vitro (McCaffrey et al. 1993). NFATc has been shown to activate IL-2 expression by a cotransfection assay in T cells (Northrop et al. 1994). Furthermore, both proteins appears to be modified by calcineurin (Jain et al. 1993; Northrop et al. 1994). In addition to NFATp and NFATc, we have isolated two new members of the human NFAT gene family. We have used these clones to examine the tissue distribution of the different NFAT genes. We have also expressed and purified the DNA binding domains of the NFAT family proteins and investigated their biochemical activities.

Results

1. Cloning of human NFAT genes

cDNA libraries were prepared from Jurkat T cells and human peripheral blood lymphocytes, and screened using a probe derived from the rel similarity region of the murine NFATp gene (McCaffrey et al. 1993). Cross-hybridizing clones were isolated, sequenced, and determined to be derived from 4 distinct genes.

One of the genes isolated in this study is related to the murine NFATp gene (McCaffrey et al. 1993), and another is identical to the NFATc gene (Northrop et al.

1994). We have isolated two classes of NFATp cDNAs which are the result of alternative splicing upstream of the rel domain. One form is similar to the cDNA reported by McCaffrey et al., while the other is alternatively spliced downstream of the rel similarity region; in particular, this form is missing an exon encoding the
5 region near the N-terminus of the protein (SEQUENCE ID NO:1, base pairs 357-867) and has a different initiating methionine (SEQUENCE ID NO:1, base pairs 880-882).

In addition to these previously identified genes, we cloned two novel members of the NFAT gene family, hereby designated as NFAT3 and NFAT4. The NFAT3 sequence was obtained from three overlapping cDNAs spanning 2880 bp, and
10 deduced to encode a protein of 902 amino acids. We obtained three classes of NFAT4 cDNAs that resulted from alternative splicing downstream of the rel homology domain. These three types of cDNAs encode proteins that vary in sequence and length at their C-terminal ends. The three forms are designated NFAT4a, NFAT4b, and NFAT4c. The positions of splice junctions in the coding regions are
15 after proline 699 in NFAT4a and after valine 700 and proline 716 in NFAT4b and NFAT4c.

All of the NFAT genes are at least 65% identical to each other within a 290 amino acid domain. This domain is related to the DNA binding and dimerization domain of the rel family of transcription factors (Nolan 1994; Northrop et al. 1994).
20 Among the different NFAT genes, the N-terminal and central portions of the rel similarity domain are more highly conserved than the C-terminus.

Aside from the strikingly similar rel domains shared by all four NFAT genes, the NFAT family members have smaller regions of sequence similarity on the amino terminal side of the rel domains. The amino terminal regions of NFAT4 and NFATc
25 have several regions of significant similarity. The two largest regions contain 23 of 41 and 24 of 45 identical amino acids between the two proteins. Both of these regions are rich in serine and proline residues. NFATp and NFAT3 also have some similarity to the other NFAT proteins in this region, although it is less extensive than that shared between NFAT4 and NFATc. The homology between NFAT3 and NFAT4 extends
30 about 25 amino acids upstream of the rel similarity region.

2. Expression patterns of the NFAT genes

On the basis of previous reports, expression of NFAT genes was expected to be restricted to lymphocytes (Shaw et al. 1988; Verweij et al. 1990; McCaffrey et al.

1993; Northrop et al. 1994). The expression of each NFAT gene was tested by Northern blot using RNA from sixteen different human tissues. For NFATp, expression of an mRNA approximately 7.5 kb was detected in almost all human tissues. The expression was slightly higher in PBLs and placenta. NFATc expression was also detected at a low level in several different tissues. The NFATc probe hybridized to two bands of approximately 2.7 and 4.5 kb. Surprisingly, the 4.5 kb NFATc transcript was strongly expressed in skeletal muscle. The 2.7 kb mRNA appears to correspond to the previously described NFATc clone (Northrop et al. 1994).

10 NFAT3 exhibited a very complicated expression pattern with at least 3 major RNA bands between 3 and 5 kb. The major sites of NFAT3 expression were observed outside the immune system. NFAT3 was highly expressed in placenta, lung, kidney, testis and ovary. In contrast, NFAT3 expression was very weak in spleen and thymus and undetectable in PBLs.

15 NFAT4 was expressed predominately as a 6.5 kb message. Like NFATc it was strongly expressed in skeletal muscle. NFAT4 also displayed relatively high expression in thymus. The probe for the NFAT4 northern contained the 3' half of the NFAT homology region as well as downstream regions from the NFAT4c class of cDNA. This probe should hybridize to all three classes of NFAT4 transcripts. Only one form is detected in the Northern blots, suggesting that the 4c class is the most abundant transcript.

These results indicate that each of the NFAT genes is expressed in a distinct tissue-specific pattern. Furthermore, none of the NFAT genes are restricted to lymphocytes.

25 3. DNA binding activity of the NFAT proteins

The rel similarity regions along with a small amount of flanking sequences of each of the four classes of NFAT proteins were expressed in E. coli. Each of the 4 proteins was well expressed and soluble. The proteins were purified to near homogeneity by DNA affinity chromatography (Kadonaga and Tjian 1986). The binding site used for purification was a high affinity NFAT site derived from the IL-4 promoter with the core binding sequence GGAAAATTTT (SEQUENCE ID NO:15) (Rooney et al. 1994).

The binding specificities of the NFAT proteins were tested on two known functional binding sites, the IL-4 promoter NFAT site and the NFAT binding site in the distal antigen response element from the IL-2 promoter (Durand et al. 1988; Shaw et al. 1988). All the proteins were able to bind the IL-4 promoter site. NFATp, 5 NFATc, and NFAT3 recognized this sequence with very similar affinity, while NFAT4 bound this sequence with lower affinity (> 10-fold) than the other three proteins in this assay. NFAT4 protein may have a different optimum binding sequence than the other NFAT proteins.

The same amounts of the four NFAT proteins were tested on the NFAT 10 binding site from the IL-2 promoter. This NFAT site (GGAAAACTG) (SEQUENCE ID NO:16) has three differences relative to the IL-4 site which make it a weaker site for all four NFAT proteins. The NFAT proteins differ in their ability to recognize this site independently. NFATp had the highest relative affinity for the IL-2 binding site, while NFATc and NFAT3 bound weakly to this site and NFAT4 binding 15 was not detectable in this assay.

The IL-2 NFAT site is part of a composite element that is adjacent to a weak AP-1 site (TGTTTCA) (Jain et al. 1992; Northrop et al. 1993). To determine if there were any differences in the ability of NFAT proteins to interact with AP-1, the four NFAT proteins were tested with AP-1 for binding to the IL-2 site. When tested alone 20 all the NFAT proteins, as well as the AP-1 proteins, bound relatively weakly to the IL-2 composite element. The combination of c-jun and fra1 with each of the four NFAT proteins resulted in highly cooperative DNA binding. In the presence of the AP-1 protein the four NFAT proteins bound to the IL-2 site with very similar affinity. In all cases, jun homodimers were not as effective as jun-fra1 heterodimers in 25 promoting cooperative binding in the gel shift assay. These results indicate that the DNA binding and protein interaction specificity of the NFAT proteins are very similar. Indeed, the interactions of the four NFAT proteins with these AP-1 proteins appear to be identical. NFAT4 did not bind independently to this site, but recognized this site with the same affinity as the other NFAT proteins in the presence of AP-1.

30 4. Transcriptional activation by the NFAT proteins

Having established that the DNA binding properties of the four NFAT proteins are quite similar, we investigated their transcriptional activation potentials. We used a transient transfection assay into Jurkat T cells to measure the ability of the NFAT

proteins to activate the IL-2 promoter. The IL-2 promoter was chosen because it is a critical regulatory target for NFAT and has at least two functional NFAT binding sites (Randak et al. 1990). Activation of this promoter by antigenic stimulation can be mimicked by treatment with phorbol esters, such as phorbol 12-myristate 13 acetate (PMA), together with ionomycin, a calcium ionophore.

Each of the four NFAT genes was transfected into Jurkat cells, and their ability to activate the IL-2 promoter was tested with various combinations of PMA and ionomycin. Treatment of the cells with PMA plus ionomycin induced strong activation by the endogenous NFAT proteins in Jurkat cells. Transfection of each of the four of the NFAT genes resulted in an additional stimulation the IL-2 promoter between 4- and 8-fold. Activation of the IL-2 promoter by each of the NFAT proteins was dependent on both PMA and ionomycin.

We also tested the ability of NFAT to activate transcription in COS and HepG2 cells using a synthetic reporter gene consisting three copies of an NFAT/AP-1 composite element. Transfection of each of the four NFAT into HepG2 cells resulted in activation of the reporter gene of at least 20-fold in the presence of PMA and ionomycin. In contrast to Jurkat cells, NFAT3 was more potent than the others in the HepG2 transfections, resulting in 140-fold activation. Another difference between the results of HepG2 and Jurkat cells is that the NFAT proteins appeared to activate transcription in the absence of PMA or calcium ionophore.

In COS cells NFAT3 produced a striking 50-fold activation that was observed independently of PMA and ionomycin treatment. NFAT3 was found to stimulate transcription in COS cells much more strongly than the other proteins.

5. NFAT proteins are active as monomers

There are many similar features of the NFAT and rel families of transcription factors. Rel proteins form homo- and heterodimers in solution, and dimerization is required for DNA binding (reviewed in Baeuerle and Henkel 1994). The C-terminal half of the rel homology domain is thought to be involved in mediating dimerization. Since the similarity between NFAT and the rel families extends throughout the 300 amino acid rel domain, and the rel domain of the NF- κ B proteins is sufficient for dimer formation, we expected that the NFAT proteins might also be function as dimers. To test this idea we determined the native masses of the NFAT proteins by gel filtration chromatography and glycerol gradient centrifugation. For these

experiments we used the rel similarity regions of NFATp and NFATc that were expressed in *E. coli* and purified by DNA affinity chromatography. The molecular weights of these proteins are 40.4 and 35.6 kD, respectively. As a control we used purified NF- κ B p50 protein that is known to exist as a stable dimer in solution

5 (Baeuerle and Baltimore 1989). The p50 protein is 45.8 kD calculated from its amino acid sequence.

On both the gel filtration column and the glycerol gradient, the NFATp and NFATc rel domains migrated at a position close to their actual molecular weight. Under the same conditions, p50 behaved as species that was larger than its monomer
10 molecular weight. The data from the gel filtration column was used to calculate the Stokes radius of each protein, and the S values were determined by glycerol gradient sedimentation. These two properties were used to calculate the apparent molecular size of the proteins (Siegel and Monty 1966; Thompson et al. 1991). The apparent molecular sizes of the NFATp and NFATc rel domains were determined to be 42 kD
15 and 32 kD respectively. These values are close to the monomer molecular weight for both NFAT proteins. As expected, p50 exhibited an apparent molecular size close to that of a dimer.

After determining that NFAT rel domains were monomers in solution, we then considered the possibility that NFAT proteins might form dimers when bound to
20 DNA. To address this question we carried out gel mobility shift assays with two different sized versions of NFATc translated in vitro (Hope and Struhl 1987). The shorter version contains the rel similarity region and a small amount of flanking residues and is referred to as NFATc-309. This construct is equivalent to the one that was expressed in *E. coli*. The larger version, NFATc-589, contains additional N-
25 terminal sequences. When expressed individually in a rabbit reticulocyte lysate both versions of NFATc were active and produced protein-DNA complexes with different mobilities. When the two different NFATc proteins were mixed by co-translation the same protein-DNA complexes were apparent and no intermediate species was detectable, as would be expected if the proteins were forming dimers on the DNA.
30 These results suggest that NFAT proteins are capable of sequence-specific DNA binding as monomers.

Methods

1. Isolation of human NFAT clones

Peripheral blood lymphocytes (PBLs) were isolated from 2 units of blood (obtained from Irwin Memorial Blood Bank, San Francisco) by fractionation on sodium metrizoate/polysaccharide (Lymphoprep, Nycomed) gradients. Jurkat T cells were grown in RPMI + 10% fetal bovine serum. Total RNA was isolated from Jurkat
5 cells or peripheral blood lymphocytes according to the Guanidinium-HCl method (Chomczynski and Sacchi 1987). Poly-A+ RNA was purified using oligo-dT magnetic beads (Promega). Random primed and oligo-dT primed libraries were prepared from both Jurkat and PBL RNA samples. The cDNA libraries were constructed in the vector Lambda ZAPII (Stratagene) according to the protocol supplied by the
10 manufacturer. The cDNA was size selected for greater than 1 kb by electrophoresis on 5% polyacrylamide gel prior to ligation. Each library contained approximately 2×10^6 recombinant clones. Each of the four libraries was screened independently under the same conditions.

The probe for the initial library screen was a 372 bp fragment derived by PCR
15 from the C-terminal half of the rel homology domain of the mouse NFATp gene. This region corresponds to amino acids 370 through 496 in the published mNFATp sequence (McCaffrey et al. 1993). The fragment was labeled by random priming and hybridized in 1M NaCl, 50 mM Tris pH 7.4, 2 mM EDTA, 10X Denhardt's, 0.05 % SDS, and 50 µg/ml salmon sperm DNA at 60°C. The filters were washed first in 2X
20 SSC, 0.1% SDS, and then in 1X SSC, 0.1% SDS at 60°C. Hybridizing clones were purified and converted into Bluescript plasmid DNA clones. The DNA sequence was determined using thermal cycle sequencing and the Applied Biosystems 373A sequencer. Approximately 50 clones were isolated from the first set of screens. Sequence analysis and cross-hybridization experiments indicated that these clones
25 were derived from 4 distinct genes. For NFAT4, additional cDNA clones were obtained from a skeletal muscle cDNA library (Stratagene). The 5' ends of the cDNA clones were obtained from a Jurkat cDNA library prepared as described above with gene specific primers for each of the NFAT genes.

2. Northern

30 The northern blots with mRNA isolated from human tissues were purchased from Clontech. DNA probes were labeled by random priming and hybridized in 5X SSPE, 10X Denhardt's, 50% formamide, 2% SDS, 100 µg/ml salmon sperm DNA at 42°C. The filters were washed in 2X SSC/0.05% SDS at room temperature, and

subsequently in 0.1X SSC/0.1% SDS at 60°C. For NFATp the probe was 1.2 kb cDNA fragment containing the entire rel similarity region of NFATp. For NFATc, the probe was a 291 nucleotide PCR fragment corresponding to the 3' end of rel similarity region (amino acids 597 to 693 (Northrop et al. 1994). For NFATc, a
5 different set of blots was hybridized with a 0.8 kb cDNA fragment located upstream of the rel domain. The two different NFATc probes produced identical results. For NFAT3, the probe was a 0.6 kb fragment located downstream of the rel similarity region corresponding to the region encoding amino acid 720 through the 3' end of the clone. For NFAT4, the probe was a 1.3 kb cDNA fragment corresponding to residue
10 549 to 963 from the 4c class of cDNAs.

3. Protein Expression and Purification

E. coli expression vectors for each NFAT protein were constructed in the T7 polymerase expression vector pT7-HMK, which has an eight amino acid heart muscle kinase (hmk) site at the N-terminus. NdeI sites were introduced by PCR using
15 mutagenic oligonucleotides in the coding regions upstream of the NFAT rel domains, and these restriction sites were subsequently used for cloning into pT7-HMK. The sizes of the different proteins (without the hmk sequences) are as follows: NFATp, 353 amino acids (the residues homologous to 185 through 537 according to McCaffrey et al. 1993); NFATc, 309 amino acids (amino acids 408 through 716
20 according to Northrop et al. 1994); NFAT3, 345 amino acids (residues 400 through 744); NFAT4, 316 amino acids (residues 393 through 708). Proteins were expressed using the T7 polymerase expression system in the strain BL21(DE3) (Studier and Moffat 1986). Expression was induced by addition of 0.4 mM IPTG, and the cultures were shaken for 4 hours at room temperature. The cells were harvested,
25 washed in PBS, resuspended in 0.4 M KCl-HEG (25 mM HEPES pH 7.9; 0.1 mM EDTA; 10% glycerol; 0.2% NP-40; 2 mM DTT, 0.2 mM PMSF, 0.2 mM sodium metabisulfite) and lysed by two cycles of freeze-thawing followed by sonication. The lysate was spun in an SS34 rotor at 10K for 10 min to remove insoluble material. NFAT proteins were purified from the soluble fractions of the extracts by DNA
30 affinity chromatography (Kadonaga and Tjian 1986). The binding site sequence for the affinity resin was from the IL-4 promoter, TACATTGGAAAATTTTATTACAC (SEQUENCE ID NO:17). The DNA was biotinylated on one strand and coupled to avidin agarose beads (Sigma) at a concentration of approximately 1 mg DNA/ml.

Approximately 10 mg of *E. coli* extracts containing the recombinant NFAT proteins were loaded on 1.5 ml DNA columns equilibrated with 0.1 M KCl-HEG. The columns were washed successively with 0.1, 0.2, and 0.4 M HEG. The specifically bound NFAT proteins were eluted with 1.0 M KCl-HEG.

- 5 Fra-1 was expressed in *E. coli* from the vector pET11 (Novagen). The protein was purified from the soluble fraction to approximately 80% homogeneity by fractionation on heparin-sepharose. c-Jun protein was expressed in *E. coli* and purified from the insoluble portion of the extract as previously described (Bohmann and Tjian, 1989). The concentrations of the purified proteins were determined by
10 comparing the intensity of coomassie staining with the staining intensity of BSA standards.

4. DNA Binding Experiments

- Electrophoretic mobility shift assays were performed with the indicated amounts of proteins in 50 mM KCl, 25 mM HEPES, 0.05 mM EDTA, 5 % glycerol, 1
15 mM DTT with 1 µg of poly(dI-dC) and 100 ng of BSA. The binding reactions and electrophoresis were carried out at room temperature. The samples were run on a 5% polyacrylamide, 0.5X TBE gel at 200 V.

5. Transfections

- The full-length coding regions for each of the NFAT genes were subcloned
20 into the RSV expression vector pREP4 (Invitrogen). The reporter plasmid was pXIL2-Luc (constructed by Jim Fraser). It contains the IL-2 promoter (-326 to +47, as in Durand et al 1988) upstream of the luciferase gene. Approximately 1×10^6 Jurkat cells were transiently transfected by lipofection (Lipofectin, Gibco/BRL). Twenty hours after transfection the cells were treated with 25 ng/ml PMA and 2 µM
25 ionomycin, and the cells were harvested 8 hours after induction. Transfection efficiencies were standardized by co-transfection of pRSV-βgal and subsequent determination of βgal activity. Each transfection contained 2 µg of expression vector, 5 µg of luciferase reporter, and 1 µg of βgal plasmid and 10 µl of lipofectin. COS-7 and HepG2 cells were transfected by a modification of the calcium phosphate method
30 (Chen and Okayama 1987). The reporter gene contained three copies of the antigen response element (-286 to -257) upstream of the herpes virus tk minimal promoter (-50 to +28) in the luciferase vector pGL2 (Promega).

6. Gel Filtration Columns and glycerol gradients

Protein samples were run on a 2.4 ml Superdex-200 column using the Pharmacia Smart system. The column was equilibrated with 0.5M KCl-HEG at a flow rate of 80 μ l/min. The elution volumes of purified NFATc, NFATp, and p50 were determined relative to those of molecular weight standards. Purified p50 was provided by Zhaodan Cao. The following molecular weight standards (10 μ g) were chromatographed on separate runs: thyroglobulin (669 kD), β -amylase (200 kD), BSA (66 kD), carbonic anhydrase (29 kD), and cytochrome c (12 kD). The elution volume (V_e) was converted to K_{av} by the equation, $K_{av} = (V_e - V_o)/V_i$, where V_o is the void volume and V_i is the included volume. The Stokes radii were determined from a plot of $(-\log K_{av})^{1/2}$ vs. the Stokes radii of the standards (Ackers 1970).

The S values were determined by glycerol gradient centrifugation. Five ml 10-30% glycerol gradients were prepared using a Beckman density gradient former. The samples were centrifuged in a SW50Ti rotor at 39,000 rpm for 40 hours. After centrifugation, 200- μ l fractions were collected and analyzed by gel electrophoresis and coomassie staining. The S values were determined by their sedimentation positions relative to the standards. Native molecular sizes were determined from the Stokes radii (a), S values (s), and the partial specific volumes (V) by the method of Siegel and Monty using the equation $M = 6\pi Nas/1-V$ (Siegel and Monty 1966, Thompson et al. 1991).

7. References cited in Experimental Section

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- 5 McCaffrey et al. (1993) *J. Biol. Chem.* **268**:3747-3752; Mouzaki and Rungger (1994) *Blood* **84**:2612-2621; Nolan (1994) *Cell* **77**:795-798; Northrop (1994) *Nature* **369**:497-502; Northrop (1993) *J. Biol. Chem.* **268**:2917-2293; Randak (1990) *EMBO J.* **9**:2529-2536; Rooney (1994) *EMBO J.* **13**:625-633; Schreiber and Crabtree (1992) *Immunol. Today* **13**:136-142; Shaw (1988) *Science* **241**:202-205; Siegel and Monty
- 10 (1966) *Biochim. Biophys. Acta* **112**:346-362; Studier and Moffat (1986) *J. Mol. Biol.* **189**:113-130; Szabo (1993) *Mol. Cell. Biol.* **13**:4793-4805; Thompson et al. (1991) *Science* **253**:762-768; Venkataraman et al. (1994) *Immunity* **1**:189-196; Verweij et al. (1990) *J. Biol. Chem.* **265**:15788-15795; Yaseen et al. (1994) *Mol. Cell. Biol.* **14**:6886-6895; and Yaseen et al. (1993) *J. Biol. Chem.* **268**:14285-14293.

15

The following examples are offered by way of illustration and not by way of limitation.

EXAMPLES

1. Protocol for hNFAT - hNFAT dependent transcription factor binding assay.
- 20 A. Reagents:
- hNFAT: 20 µg/ml in PBS.
 - Blocking buffer: 5% BSA, 0.5% Tween 20 in PBS; 1 hr. RT.
 - Assay Buffer: 100 mM KCl, 20 mM HEPES pH 7.6, 0.25 mM EDTA, 1% glycerol, 0.5 % NP-40, 50 mM BME, 1 mg/ml BSA, cocktail of protease inhibitors.
- 25 - ³²P hNFAT 10x stock: 10^{-8} - 10^{-6} M "cold" hNFAT homolog supplemented with 200,000-250,000 cpm of labeled hNFAT homolog (Beckman counter). Place in the 4 °C microfridge during screening.
- Protease inhibitor cocktail (1000X): 10 mg Trypsin Inhibitor (BMB # 109894), 10 mg Aprotinin (BMB # 236624), 25 mg Benzamidine (Sigma # B-6506),
- 30 25 mg Leupeptin (BMB # 1017128), 10 mg APMSF (BMB # 917575), and 2mM NaVO₃ (Sigma # S-6508) in 10 ml of PBS.
- B. Preparation of assay plates:

- Coat with 120 µl of stock NF-AT per well overnight at 4 °C.
 - Wash 2X with 200 µl PBS.
 - Block with 150 µl of blocking buffer.
 - Wash 2X with 200 µl PBS.
- 5 C. Assay:
- Add 80 µl assay buffer/well.
 - Add 10 µl compound or extract.
 - Add 10 µl ³³P-NFAT (20,000-25,000 cpm/0.3 pmoles/well = 3x10⁻⁹ M final concentration).
- 10
- Shake at 25C for 15 min.
 - Incubate additional 45 min. at 25C.
 - Stop the reaction by washing 4X with 200 µl PBS.
 - Add 150 µl scintillation cocktail.
 - Count in Topcount.
- 15 D. Controls for all assays (located on each plate):
- a. Non-specific binding (no hNFAT added)
 - b. cold hNFAT at 80% inhibition.
2. Protocol for hNFAT - AP1 dependent transcription factor binding assay.
- 20 A. Reagents:
- fos-jun heterodimers (junB and fra1): 20 µg/ml in PBS.
 - Blocking buffer: 5% BSA, 0.5% Tween 20 in PBS; 1 hr, RT.
 - Assay Buffer: 100 mM KCl, 20 mM HEPES pH 7.6, 0.25 mM EDTA, 1% glycerol, 0.5 % NP-40, 50 mM BME, 1 mg/ml BSA, cocktail of protease inhibitors.
- 25
- ³³P hNFAT 10x stock: 10⁻⁸ - 10⁻⁶ M "cold" hNFAT homolog supplemented with 200,000-250,000 cpm of labeled hNFAT homolog (Beckman counter). Place in the 4 °C microfridge during screening.
 - Protease inhibitor cocktail (1000X): 10 mg Trypsin Inhibitor (BMB # 109894), 10 mg Aprotinin (BMB # 236624), 25 mg Benzamidine (Sigma # B-6506), 25 mg Leupeptin (BMB # 1017128), 10 mg APMSF (BMB # 917575), and 2mM NaVO₃ (Sigma # S-6508) in 10 ml of PBS.
- 30
- B. Preparation of assay plates:
- Coat with 120 µl of stock fos-jun heterodimers per well overnight at 4 °C.

- Wash 2X with 200 µl PBS.
- Block with 150 µl of blocking buffer.
- Wash 2X with 200 µl PBS.
- C. Assay:
 - 5 - Add 80 µl assay buffer/well.
 - Add 10 µl compound or extract.
 - Add 10 µl ^{33}P -NFAT (20,000-25,000 cpm/0.3 pmoles/well = 3×10^{-9} M final concentration).
 - Shake at 25C for 15 min.
 - 10 - Incubate additional 45 min. at 25C.
 - Stop the reaction by washing 4X with 200 µl PBS.
 - Add 150 µl scintillation cocktail.
 - Count in Topcount.
- D. Controls for all assays (located on each plate):
 - 15 a. Non-specific binding (no hNFAT added)
 - b. cold hNFAT at 80% inhibition.
- 3. Protocol for hNFAT-fos-jun dependent transcription factor - DNA binding assay.
- 20 A. Reagents:
 - Neutralite Avidin: 20 µg/ml in PBS.
 - Blocking buffer: 5% BSA, 0.5% Tween 20 in PBS; 1 hr, RT.
 - Assay Buffer: 100 mM KCl, 20 mM HEPES pH 7.6, 0.25 mM EDTA, 1% glycerol, 0.5 % NP-40, 50 mM BME, 1 mg/ml BSA, cocktail of protease inhibitors.
 - 25 - ^{33}P hNFAT 10x stock: 10^{-6} - 10^{-8} M "cold" hNFAT homolog supplemented with 200,000-250,000 cpm of labeled hNFAT homolog (Beckman counter) and 10^{-6} - 10^{-8} M fos-jun heterodimers. Place in the 4 °C microfridge during screening.
 - Protease inhibitor cocktail (1000X): 10 mg Trypsin Inhibitor (BMB # 109894), 10 mg Aprotinin (BMB # 236624), 25 mg Benzamidine (Sigma # B-6506),
 - 30 25 mg Leupeptin (BMB # 1017128), 10 mg APMSF (BMB # 917575), and 2mM NaVO_3 (Sigma # S-6508) in 10 ml of PBS.

- Oligonucleotide stock: (specific biotinylated). Biotinylated oligo at 17 pmole/ μ l, AP1-NFAT site: (BIOTIN)-GG AGG AAA AAC TGT TTC ATA CAG AAG GCG T (SEQUENCE ID NO:18)

B. Preparation of assay plates:

- 5
- Coat with 120 μ l of stock N-Avidin per well overnight at 4 °C.
 - Wash 2X with 200 μ l PBS.
 - Block with 150 μ l of blocking buffer.
 - Wash 2X with 200 μ l PBS.

C. Assay:

- 10
- Add 40 μ l assay buffer/well.
 - Add 10 μ l compound or extract.
 - Add 10 μ l 33 P-NFAT (20,000-25,000 cpm/0.1-10 pmoles/well = 10^{-9} - 10^{-7} M final concentration).

- Shake at 25C for 15 min.

- 15
- Incubate additional 45 min. at 25C.
 - Add 40 μ l oligo mixture (1.0 pmoles/40 μ l in assay buffer with 1 ng of ss-DNA)

- Incubate 1 hr at RT.

- Stop the reaction by washing 4X with 200 μ l PBS.

- 20
- Add 150 μ l scintillation cocktail.
 - Count in Topcount.

D. Controls for all assays (located on each plate):

- a. Non-specific binding (no oligo added)
- b. Specific soluble oligo at 80% inhibition.

25

All publications and patent applications cited in this specification are herein incorporated by reference as if each individual publication or patent application were specifically and individually indicated to be incorporated by reference. Although the foregoing invention has been described in some detail by way of illustration and

30 example for purposes of clarity of understanding, it will be readily apparent to those of ordinary skill in the art in light of the teachings of this invention that certain changes and modifications may be made thereto without departing from the spirit or scope of the appended claims.

SEQUENCE LISTING

(1) GENERAL INFORMATION:

- (i) APPLICANT: HOEY, Timothy
- (ii) TITLE OF INVENTION: NUCLEAR FACTORS AND BINDING ASSAY
- (iii) NUMBER OF SEQUENCES: 18
- (iv) CORRESPONDENCE ADDRESS:
 - (A) ADDRESSEE: FLEHR, HOHBACH, TEST, ALBRITTON & HERBERT
 - (B) STREET: 4 Embarcadero Center, Suite 3400
 - (C) CITY: San Francisco
 - (D) STATE: California
 - (E) COUNTRY: USA
 - (F) ZIP: 94111
- (v) COMPUTER READABLE FORM:
 - (A) MEDIUM TYPE: Floppy disk
 - (B) COMPUTER: IBM PC compatible
 - (C) OPERATING SYSTEM: PC-DOS/MS-DOS
 - (D) SOFTWARE: PatentIn Release #1.0, Version #1.30
- (vi) CURRENT APPLICATION DATA:
 - (A) APPLICATION NUMBER: US
 - (B) FILING DATE:
 - (C) CLASSIFICATION:
- (viii) ATTORNEY/AGENT INFORMATION:
 - (A) NAME: Osman, Richard A
 - (B) REGISTRATION NUMBER: 36,627
 - (C) REFERENCE/DOCKET NUMBER: A-59450-1/RAO
- (ix) TELECOMMUNICATION INFORMATION:
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 - (B) TELEFAX: (415) 494-8771
 - (C) TELEX: 210 277299

(2) INFORMATION FOR SEQ ID NO:1:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 3478 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

- (ix) FEATURE:
 - (A) NAME/KEY: CDS
 - (B) LOCATION: 223..2987

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

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ACCCGCTGCG CCAGGCGCGC CGTCCCCGGA CCCGGCGTGC GTCCCTACGA GGAAAGGGAC      120
CCCGCCGCTC GAGCCGCCTC CGCCAGCCCC ACTGCGAGGG GTCCCAGAGC CAGCCGCGCC      180
CGCCCTCGCC CCCGGCCCCG CAGCCTTCCC GCCCTGCGCG CC ATG AAC GCC CCC      234
                                     Met Asn Ala Pro

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Glu Arg Gln Pro Gln Pro Asp Gly Gly Asp Ala Pro Gly His Glu Pro	
5 10 15 20	
GGG GGC AGC CCC CAA GAC GAG CTT GAC TTC TCC ATC CTC TTC GAC TAT	330
Gly Gly Ser Pro Gln Asp Glu Leu Asp Phe Ser Ile Leu Phe Asp Tyr	
25 30 35	
GAG TAT TTG AAT CCG AAC GAA GAA GAG CCG AAT GCA CAT AAG GTC GCC	378
Glu Tyr Leu Asn Pro Asn Glu Glu Glu Pro Asn Ala His Lys Val Ala	
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AGC CCA CCC TCC GGA CCC GCA TAC CCC GAT GAT GTC CTG GAC TAT GGC	426
Ser Pro Pro Ser Gly Pro Ala Tyr Pro Asp Asp Val Leu Asp Tyr Gly	
55 60 65	
CTC AAG CCA TAC AGC CCC CTT GCT AGT CTC TCT GGC GAG CCC CCC GGC	474
Leu Lys Pro Tyr Ser Pro Leu Ala Ser Leu Ser Gly Glu Pro Pro Gly	
70 75 80	
CGA TTC GGA GAG CCG GAT AGG GTA GGG CCG CAG AAG TTT CTG AGC GCG	522
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85 90 95 100	
GCC AAG CCA GCA GGG GCC TCG GGC CTG AGC CCT CGG ATC GAG ATC ACT	570
Ala Lys Pro Ala Gly Ala Ser Gly Leu Ser Pro Arg Ile Glu Ile Thr	
105 110 115	
CCG TCC CAC GAA CTG ATC CAG GCA GTG GGG CCC CTC CGC ATG AGA GAC	618
Pro Ser His Glu Leu Ile Gln Ala Val Gly Pro Leu Arg Met Arg Asp	
120 125 130	
GCG GGC CTC CTG GTG GAG CAG CCG CCC CTG GCC GGG GTG GCC GCC AGC	666
Ala Gly Leu Leu Val Glu Gln Pro Pro Leu Ala Gly Val Ala Ala Ser	
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CCG AGG TTC ACC CTG CCC GTG CCC GGC TTC GAG GGC TAC CGC GAG CCG	714
Pro Arg Phe Thr Leu Pro Val Pro Gly Phe Glu Gly Tyr Arg Glu Pro	
150 155 160	
CTT TGC TTG AGC CCC GCT AGC AGC GGC TCC TCT GCC AGC TTC ATT TCT	762
Leu Cys Leu Ser Pro Ala Ser Ser Gly Ser Ser Ala Ser Phe Ile Ser	
165 170 175 180	
GAC ACC TTC TCC CCC TAC ACC TCG CCC TGC GTC TCG CCC AAT AAC GGC	810
Asp Thr Phe Ser Pro Tyr Thr Ser Pro Cys Val Ser Pro Asn Asn Gly	
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Asp Ser Cys Leu Gly Arg His Ser Pro Val Pro Arg Pro Ala Ser Arg	
230 235 240	
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Ser Ser Ser Pro Gly Ala Lys Arg Arg His Ser Cys Ala Glu Ala Leu	
245 250 255 260	
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Val Ala Leu Pro Pro Gly Ala Ser Pro Gln Arg Ser Arg Ser Pro Ser	
265 270 275	
CCG CAG CCC TCA TCT CAC GTG GCA CCC CAG GAC CAC GGC TCC CCG GCT	1098

Pro	Gln	Pro	Ser	Ser	His	Val	Ala	Pro	Gln	Asp	His	Gly	Ser	Pro	Ala		
			280					285					290				
GGG	TAC	CCC	CCT	GTG	GCT	GGC	TCT	GCC	GTG	ATC	ATG	GAT	GCC	CTG	AAC	1146	
Gly	Tyr	Pro	Pro	Val	Ala	Gly	Ser	Ala	Val	Ile	Met	Asp	Ala	Leu	Asn		
		295					300					305					
AGC	CTC	GCC	ACG	GAC	TCG	CCT	TGT	GGG	ATC	CCC	CCC	AAG	ATG	TGG	AAG	1194	
Ser	Leu	Ala	Thr	Asp	Ser	Pro	Cys	Gly	Ile	Pro	Pro	Lys	Met	Trp	Lys		
	310					315					320						
ACC	AGC	CCT	GAC	CCC	TCG	CCG	GTG	TCT	GCC	GCC	CCA	TCC	AAG	GCC	GGC	1242	
Thr	Ser	Pro	Asp	Pro	Ser	Pro	Val	Ser	Ala	Ala	Pro	Ser	Lys	Ala	Gly		
	325				330				335						340		
CTG	CCT	CGC	CAC	ATC	TAC	CCG	GCC	GTG	GAG	TTC	CTG	GGG	CCC	TGC	GAG	1290	
Leu	Pro	Arg	His	Ile	Tyr	Pro	Ala	Val	Glu	Phe	Leu	Gly	Pro	Cys	Glu		
			345					350						355			
CAG	GGC	GAG	AGG	AGA	AAC	TCG	GCT	CCA	GAA	TCC	ATC	CTG	CTG	GTT	CCG	1338	
Gln	Gly	Glu	Arg	Arg	Asn	Ser	Ala	Pro	Glu	Ser	Ile	Leu	Leu	Val	Pro		
			360					365					370				
CCC	ACT	TGG	CCC	AAG	CCG	CTG	GTG	CCT	GCC	ATT	CCC	ATC	TGC	AGC	ATC	1386	
Pro	Thr	Trp	Pro	Lys	Pro	Leu	Val	Pro	Ala	Ile	Pro	Ile	Cys	Ser	Ile		
		375				380						385					
CCA	GTG	ACT	GCA	TCC	CTC	CCT	CCA	CTT	GAG	TGG	CCG	CTG	TCC	AGT	CAG	1434	
Pro	Val	Thr	Ala	Ser	Leu	Pro	Pro	Leu	Glu	Trp	Pro	Leu	Ser	Ser	Gln		
	390					395					400						
TCA	GGC	TCT	TAC	GAG	CTG	CGG	ATC	GAG	GTG	CAG	CCC	AAG	CCA	CAT	CAC	1482	
Ser	Gly	Ser	Tyr	Glu	Leu	Arg	Ile	Glu	Val	Gln	Pro	Lys	Pro	His	His		
	405			410				415						420			
CGG	GCC	CAC	TAT	GAG	ACA	GAA	GGC	AGC	CGA	GGG	GCT	GTC	AAA	GCT	CCA	1530	
Arg	Ala	His	Tyr	Glu	Thr	Glu	Gly	Ser	Arg	Gly	Ala	Val	Lys	Ala	Pro		
			425					430					435				
ACT	GGA	GGC	CAC	CCT	GTG	GTT	CAG	CTC	CAT	GGC	TAC	ATG	GAA	AAC	AAG	1578	
Thr	Gly	Gly	His	Pro	Val	Val	Gln	Leu	His	Gly	Tyr	Met	Glu	Asn	Lys		
			440				445						450				
CCT	CTG	GGA	CTT	CAG	ATC	TTC	ATT	GGG	ACA	GCT	GAT	GAG	CGG	ATC	CTT	1626	
Pro	Leu	Gly	Leu	Gln	Ile	Phe	Ile	Gly	Thr	Ala	Asp	Glu	Arg	Ile	Leu		
		455				460						465					
AAG	CCG	CAC	GCC	TTC	TAC	CAG	GTG	CAC	CGA	ATC	ACG	GGG	AAA	ACT	GTC	1674	
Lys	Pro	His	Ala	Phe	Tyr	Gln	Val	His	Arg	Ile	Thr	Gly	Lys	Thr	Val		
		470				475					480						
ACC	ACC	ACC	AGC	TAT	GAG	AAG	ATA	GTG	GGC	AAC	ACC	AAA	GTC	CTG	GAG	1722	
Thr	Thr	Thr	Ser	Tyr	Glu	Lys	Ile	Val	Gly	Asn	Thr	Lys	Val	Leu	Glu		
					490				495					500			
ATA	CCC	TTG	GAG	CCC	AAA	AAC	AAC	ATG	AGG	GCA	ACC	ATC	GAC	TGT	GCG	1770	
Ile	Pro	Leu	Glu	Pro	Lys	Asn	Asn	Met	Arg	Ala	Thr	Ile	Asp	Cys	Ala		
				505					510					515			
GGG	ATC	TTG	AAG	CTT	AGA	AAC	GCC	GAC	ATT	GAG	CTG	CGG	AAA	GGC	GAG	1818	
Gly	Ile	Leu	Lys	Leu	Arg	Asn	Ala	Asp	Ile	Glu	Leu	Arg	Lys	Gly	Glu		
			520					525					530				
ACG	GAC	ATT	GGA	AGA	AAG	AAC	ACG	CGG	GTG	AGA	CTG	GTT	TTC	CGA	GTT	1866	
Thr	Asp	Ile	Gly	Arg	Lys	Asn	Thr	Arg	Val	Arg	Leu	Val	Phe	Arg	Val		
		535					540					545					
CAC	ATC	CCA	GAG	TCC	AGT	GGC	AGA	ATC	GTC	TCT	TTA	CAG	ACT	GCA	TCT	1914	
His	Ile	Pro	Glu	Ser	Ser	Gly	Arg	Ile	Val	Ser	Leu	Gln	Thr	Ala	Ser		

550	555	560	
AAC Asn 565	CCC Pro 570	ATC Ile 575	1962
GAG Glu 580	TGC Cys 585	TCC Ser 590	
CAG Gln 595	CGA Arg 600	TCT Ser 605	
GCT Ala 610	CAC His 615	GAG Glu 620	
CTG Leu 625	CCC Pro 630	ATG Met 635	
GTT Val 640			
GAA Glu 645	AGA Arg 650	CAA Gln 655	2010
GAC Asp 660	ACA Thr 665	GAC Asp 670	
AGC Ser 675	TGC Cys 680	CTG Leu 685	
GTC Val 690	TAT Tyr 695	GGC Gly 700	
GGC Gly 705	CAG Gln 710	CAA Gln 715	
ATG Met 720			
ATC Ile 725	CTC Leu 730	ACG Thr 735	2058
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TTT Phe 755	ACA Thr 760	TCC Ser 765	
GAG Glu 770	TCC Ser 775	AAA Lys 780	
GTT Val 785	GTG Val 790	TTT Phe 795	
ACT Thr 800			
GAG Glu 805	AAG Lys 810	ACC Thr 815	2106
ACA Thr 820	GAT Asp 825	GGA Gly 830	
CAG Gln 835	CAA Gln 840	ATT Ile 845	
TGG Trp 850	GAG Glu 855	ATG Met 860	
GAA Glu 865	GCC Ala 870	ACG Thr 875	
GTG Val 880			
GAT Asp 885	AAG Lys 890	GAC Asp 895	2154
AAG Lys 900	GAC Asp 905	AGC Ser 910	
CAG Gln 915	CCC Pro 920	AAC Asn 925	
ATG Met 930	ATT Ile 935	TGG Trp 940	
CTT Leu 945	TTT Phe 950	GTT Val 955	
GAG Glu 960	ATC Ile 965	CCT Pro 970	
GAA Glu 975			
CGG Arg 980	AAC Asn 985	AAG Lys 990	2202
AAG Lys 995	CAT His 1000	ATC Ile 1005	
CGC Arg 1010	ACA Thr 1015	CCT Pro 1020	
GTA Val 1025	AAA Lys 1030	GTG Val 1035	
AAA Lys 1040	GTG Val 1045	AAC Asn 1050	
TTC Phe 1055	TAC Tyr 1060	GTC Val 1065	
ATC Ile 1070	AAT Asn 1075	GGG Gly 1080	2250
AGG Arg 1085	AAA Lys 1090	CGA Arg 1095	
AGT Ser 1100	CAG Gln 1105	CCT Pro 1110	
CAG Gln 1115	CAC His 1120	TTT Phe 1125	
ACC Thr 1130	TAC Tyr 1135	CAC His 1140	
CCA Pro 1145	GTC Val 1150	CCA Pro 1155	2298
GCC Ala 1160	ATC Ile 1165	AAG Lys 1170	
ACG Thr 1175	GAG Glu 1180	CCC Pro 1185	
ACG Thr 1190	GAT Asp 1195	GAA Glu 1200	
TAT Tyr 1205	GAC Asp 1210	CCC Pro 1215	
ACT Thr 1220			
CTG Leu 1225	ATC Ile 1230	TGC Cys 1235	2346
AGC Ser 1240	CCC Pro 1245	ACC Thr 1250	
CAT His 1255	GGA Gly 1260	GGC Gly 1265	
GGA Gly 1270	GGC Gly 1275	CTG Leu 1280	
GGG Gly 1285	AGC Ser 1290	CAG Gln 1295	
CCT Pro 1300	TAC Tyr 1305	TAC Tyr 1310	
CCC Pro 1315	CAG Gln 1320	CCC Pro 1325	2394
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GCC Ala 1375			
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CCC Pro 1395	GGG Gly 1400	CTC Leu 1405	
TCA Ser 1410	TCC Ser 1415	CTC Leu 1420	
TCC Ser 1425	TGC Cys 1430	CTC Leu 1435	
CTC Leu 1440	CTC Leu 1445	GTG Val 1450	
GCC Ala 1455	ACC Thr 1460	GCC Ala 1465	
ACC Thr 1470			
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AAC Asn 1505	CCA Pro 1510	GCG Ala 1515	
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AAG Lys 1565			
AGC Ser 1570	CTG Leu 1575	CTG Leu 1580	2538
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CTG Leu 1600	CTG Leu 1605	GGC Gly 1610	
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TCT Ser 1695	GTG Val 1700	CTG Leu 1705	
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TCA Ser 1760	GCC Ala 1765	CTG Leu 1770	
CTC Leu 1775	CTC Leu 1780	CAC His 1785	
CCC Pro 1790	CCC Pro 1795	1CT Ser 1800	
CCG Arg 1805	ACC Thr 1810	CCG Arg 1815	
ACC Thr 1820	AAC Asn 1825	CAG Gln 1830	2682
CAG Gln 1835	CAG Gln 1840	CAG Gln 1845	
CTG Leu 1850			
CGC Arg 1855	TGC Cys 1860	GGA Gly 1865	2730
AGC Ser 1870	CAC His 1875	CAG Gln 1880	
GAG Glu 1885	TTC Phe 1890	CAG Gln 1895	
CAC His 1900	CAG Gln 1905	CAC His 1910	
ATC Ile 1915	ATG Met 1920	TAC Tyr 1925	
TGC Cys 1930	GAG Glu 1935	AAT Asn 1940	

TTC GCA CCA GGC ACC ACC AGA CCT GGC CCG CCC CCG GTC AGT CAA GGT	2778
Phe Ala Pro Gly Thr Thr Arg Pro Gly Pro Pro Pro Val Ser Gln Gly	
840 845 850	
CAG AGG CTG AGC CCG GGT TCC TAC CCC ACA GTC ATT CAG CAG CAG AAT	2826
Gln Arg Leu Ser Pro Gly Ser Tyr Pro Thr Val Ile Gln Gln Gln Asn	
855 860 865	
GCC ACG AGC CAA AGA GCC GCC AAA AAC GGA CCC CCG GTC AGT GAC CAA	2874
Ala Thr Ser Gln Arg Ala Ala Lys Asn Gly Pro Pro Val Ser Asp Gln	
870 875 880	
AAG GAA GTA TTA CCT GCG GGG GTG ACC ATT AAA CAG GAG CAG AAC TTG	2922
Lys Glu Val Leu Pro Ala Gly Val Thr Ile Lys Gln Glu Gln Asn Leu	
885 890 895 900	
GAC CAG ACC TAC TTG GAT GAT GAG CTG ATA GAC ACA CAC CTT AGC TGG	2970
Asp Gln Thr Tyr Leu Asp Asp Glu Leu Ile Asp Thr His Leu Ser Trp	
905 910 915	
ATA CAA AAC ATA TTA TG AAACAGAATG ACTGTGATCT TTGATCCGAG	3017
Ile Gln Asn Ile Leu	
920	
AAATCAAAGT TAAAGTTAAT GAAATTATCA GGAAGGAGTT TTCAGGACCT CCTGCCAGAA	3077
ATCAGACGTA AAAGAAGCCA TTATAGCAAG ACACCTTCTG TATCTGACCC CTCGGAGCCC	3137
TCCACAGCCC CTCACCTTCT GTCTCCTTTC ATGTTTCATCT CCCAGCCCGG AGTCCACACG	3197
CGGATCAATG TATGGGCACT AAGCGGACTC TCACTTAAGG AGCTCGCCAC CTCCTCTAA	3257
ACACCAGAGA GAACTCTTCT TTTCGGTTTA TGTTTTAAAT CCCAGAGAGC ATCCTGGTTG	3317
ATCTTAATGG TGTTCCGTCC AAATAGTAAG CACCTGCTGA CCAAAGCAC ATTCTACATG	3377
AGACAGGACA CTGGAATCTT CCTGAGAACA GAGTGACTGG AGCTTGGGGG GATGGACGGG	3437
GGACAGAAGA TGTGGGCACT GTGATTAAAC CCCAGCCCTT G	3478

(2) INFORMATION FOR SEQ ID NO:2:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 921 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

Met Asn Ala Pro Glu Arg Gln Pro Gln Pro Asp Gly Gly Asp Ala Pro
 1 5 10 15

Gly His Glu Pro Gly Gly Ser Pro Gln Asp Glu Leu Asp Phe Ser Ile
 20 25 30

Leu Phe Asp Tyr Glu Tyr Leu Asn Pro Asn Glu Glu Glu Pro Asn Ala
 35 40 45

His Lys Val Ala Ser Pro Pro Ser Gly Pro Ala Tyr Pro Asp Asp Val
 50 55 60

Leu Asp Tyr Gly Leu Lys Pro Tyr Ser Pro Leu Ala Ser Leu Ser Gly
 65 70 75 80

Glu Pro Pro Gly Arg Phe Gly Glu Pro Asp Arg Val Gly Pro Gln Lys
 85 90 95

Phe Leu Ser Ala Ala Lys Pro Ala Gly Ala Ser Gly Leu Ser Pro Arg
 100 105 110
 Ile Glu Ile Thr Pro Ser His Glu Leu Ile Gln Ala Val Gly Pro Leu
 115 120 125
 Arg Met Arg Asp Ala Gly Leu Leu Val Glu Gln Pro Pro Leu Ala Gly
 130 135 140
 Val Ala Ala Ser Pro Arg Phe Thr Leu Pro Val Pro Gly Phe Glu Gly
 145 150 155 160
 Tyr Arg Glu Pro Leu Cys Leu Ser Pro Ala Ser Ser Gly Ser Ser Ala
 165 170 175
 Ser Phe Ile Ser Asp Thr Phe Ser Pro Tyr Thr Ser Pro Cys Val Ser
 180 185 190
 Pro Asn Asn Gly Gly Pro Asp Asp Leu Cys Pro Gln Phe Gln Asn Ile
 195 200 205
 Pro Ala His Tyr Ser Pro Arg Thr Ser Pro Ile Met Ser Pro Arg Thr
 210 215 220
 Ser Leu Ala Glu Asp Ser Cys Leu Gly Arg His Ser Pro Val Pro Arg
 225 230 235 240
 Pro Ala Ser Arg Ser Ser Ser Pro Gly Ala Lys Arg Arg His Ser Cys
 245 250 255
 Ala Glu Ala Leu Val Ala Leu Pro Pro Gly Ala Ser Pro Gln Arg Ser
 260 265 270
 Arg Ser Pro Ser Pro Gln Pro Ser Ser His Val Ala Pro Gln Asp His
 275 280 285
 Gly Ser Pro Ala Gly Tyr Pro Pro Val Ala Gly Ser Ala Val Ile Met
 290 295 300
 Asp Ala Leu Asn Ser Leu Ala Thr Asp Ser Pro Cys Gly Ile Pro Pro
 305 310 315 320
 Lys Met Trp Lys Thr Ser Pro Asp Pro Ser Pro Val Ser Ala Ala Pro
 325 330 335
 Ser Lys Ala Gly Leu Pro Arg His Ile Tyr Pro Ala Val Glu Phe Leu
 340 345 350
 Gly Pro Cys Glu Gln Gly Glu Arg Arg Asn Ser Ala Pro Glu Ser Ile
 355 360 365
 Leu Leu Val Pro Pro Thr Trp Pro Lys Pro Leu Val Pro Ala Ile Pro
 370 375 380
 Ile Cys Ser Ile Pro Val Thr Ala Ser Leu Pro Pro Leu Glu Trp Pro
 385 390 395 400
 Leu Ser Ser Gln Ser Gly Ser Tyr Glu Leu Arg Ile Glu Val Gln Pro
 405 410 415
 Lys Pro His His Arg Ala His Tyr Glu Thr Glu Gly Ser Arg Gly Ala
 420 425 430
 Val Lys Ala Pro Thr Gly Gly His Pro Val Val Gln Leu His Gly Tyr
 435 440 445
 Met Glu Asn Lys Pro Leu Gly Leu Gln Ile Phe Ile Gly Thr Ala Asp
 450 455 460

Glu Arg Ile Leu Lys Pro His Ala Phe Tyr Gln Val His Arg Ile Thr
 465 470 475 480
 Gly Lys Thr Val Thr Thr Thr Ser Tyr Glu Lys Ile Val Gly Asn Thr
 485 490 495
 Lys Val Leu Glu Ile Pro Leu Glu Pro Lys Asn Asn Met Arg Ala Thr
 500 505 510
 Ile Asp Cys Ala Gly Ile Leu Lys Leu Arg Asn Ala Asp Ile Glu Leu
 515 520 525
 Arg Lys Gly Glu Thr Asp Ile Gly Arg Lys Asn Thr Arg Val Arg Leu
 530 535 540
 Val Phe Arg Val His Ile Pro Glu Ser Ser Gly Arg Ile Val Ser Leu
 545 550 555 560
 Gln Thr Ala Ser Asn Pro Ile Glu Cys Ser Gln Arg Ser Ala His Glu
 565 570 575
 Leu Pro Met Val Glu Arg Gln Asp Thr Asp Ser Cys Leu Val Tyr Gly
 580 585 590
 Gly Gln Gln Met Ile Leu Thr Gly Gln Asn Phe Thr Ser Glu Ser Lys
 595 600 605
 Val Val Phe Thr Glu Lys Thr Thr Asp Gly Gln Gln Ile Trp Glu Met
 610 615 620
 Glu Ala Thr Val Asp Lys Asp Lys Ser Gln Pro Asn Met Leu Phe Val
 625 630 635 640
 Glu Ile Pro Glu Tyr Arg Asn Lys His Ile Arg Thr Pro Val Lys Val
 645 650 655
 Asn Phe Tyr Val Ile Asn Gly Lys Arg Lys Arg Ser Gln Pro Gln His
 660 665 670
 Phe Thr Tyr His Pro Val Pro Ala Ile Lys Thr Glu Pro Thr Asp Glu
 675 680 685
 Tyr Asp Pro Thr Leu Ile Cys Ser Pro Thr His Gly Gly Leu Gly Ser
 690 695 700
 Gln Pro Tyr Tyr Pro Gln His Pro Met Val Ala Glu Ser Pro Ser Cys
 705 710 715 720
 Leu Val Ala Thr Met Ala Pro Cys Gln Gln Phe Arg Thr Gly Leu Ser
 725 730 735
 Ser Pro Asp Ala Arg Tyr Gln Gln Gln Asn Pro Ala Ala Val Leu Tyr
 740 745 750
 Gln Arg Ser Lys Ser Leu Ser Pro Ser Leu Leu Gly Tyr Gln Gln Pro
 755 760 765
 Ala Leu Met Ala Ala Pro Leu Ser Leu Ala Asp Ala His Arg Ser Val
 770 775 780
 Leu Val His Ala Gly Ser Gln Gly Gln Ser Ser Ala Leu Leu His Pro
 785 790 795 800
 Ser Pro Thr Asn Gln Gln Ala Ser Pro Val Ile His Tyr Ser Pro Thr
 805 810 815
 Asn Gln Gln Leu Arg Cys Gly Ser His Gln Glu Phe Gln His Ile Met
 820 825 830

Tyr Cys Glu Asn Phe Ala Pro Gly Thr Thr Arg Pro Gly Pro Pro Pro
 835 840 845
 Val Ser Gln Gly Gln Arg Leu Ser Pro Gly Ser Tyr Pro Thr Val Ile
 850 855 860
 Gln Gln Gln Asn Ala Thr Ser Gln Arg Ala Ala Lys Asn Gly Pro Pro
 865 870 875 880
 Val Ser Asp Gln Lys Glu Val Leu Pro Ala Gly Val Thr Ile Lys Gln
 885 890 895
 Glu Gln Asn Leu Asp Gln Thr Tyr Leu Asp Asp Glu Leu Ile Asp Thr
 900 905 910
 His Leu Ser Trp Ile Gln Asn Ile Leu
 915 920

(2) INFORMATION FOR SEQ ID NO:3:

- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 2743 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 240..2390

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

GAATTCCGCA GGGCGCGGGC ACCGGGGCGC GGGCAGGGCT CGGAGCCACC GCGCAGGTCC	60
TAGGGCCGCG GCCGGGCCCC GCCACGCGCG CACACGCCCC TCGATGACTT TCCTCCGGGG	120
CGCGCGGCGC TGAGCCCGGG GCGAGGGCTG TCTTCCCGGA GACCCGACCC CGGCAGCGCG	180
GGGCGGCCAC TTCTCCTGTG CCTCCGCCCC CTGCTCCACT CCCCGCCGCC GCCGCGCG	239
ATG CCA AGC ACC AGC TTT CCA GTC CCT TCC AAG TTT CCA CTT GGC CCT	287
Met Pro Ser Thr Ser Phe Pro Val Pro Ser Lys Phe Pro Leu Gly Pro	
925 930 935	
GCG GCT GCG GTC TTC GGG AGA GGA GAA ACT TTG GGG CCC GCG CCG CGC	335
Ala Ala Ala Val Phe Gly Arg Gly Glu Thr Leu Gly Pro Ala Pro Arg	
940 945 950	
GCC GGC GGC ACC ATG AAG TCA GCG GAG GAA GAA CAC TAT GGC TAT GCA	383
Ala Gly Gly Thr Met Lys Ser Ala Glu Glu Glu His Tyr Gly Tyr Ala	
955 960 965	
TCC TCC AAC GTC AGC CCC GCC CTG CCG CTC CCC ACG GCG CAC TCC ACC	431
Ser Ser Asn Val Ser Pro Ala Leu Pro Leu Pro Thr Ala His Ser Thr	
970 975 980 985	
CTG CCG GCC CCG TGC CAC AAC CTT CAG ACC TCC ACA CCG GGC ATC ATC	479
Leu Pro Ala Pro Cys His Asn Leu Gln Thr Ser Thr Pro Gly Ile Ile	
990 995 1000	
CCG CCG GCG GAT CAC CCC TCG GGG TAC GGA GCA GCT TTG GAC GGT GGG	527
Pro Pro Ala Asp His Pro Ser Gly Tyr Gly Ala Ala Leu Asp Gly Gly	
1005 1010 1015	
CCC GCG GGC TAC TTC CTC TCC TCC GGC CAC ACC AGG CCT GAT GGG GCC	575

Pro	Ala	Gly	Tyr	Phe	Leu	Ser	Ser	Gly	His	Thr	Arg	Pro	Asp	Gly	Ala		
	1020						1025					1030					
CCT	GCC	CTG	GAG	AGT	CCT	CGC	ATC	GAG	ATA	ACC	TCG	TGC	TTG	GGC	CTG	623	
Pro	Ala	Leu	Glu	Ser	Pro	Arg	Ile	Glu	Ile	Thr	Ser	Cys	Leu	Gly	Leu		
	1035					1040					1045						
TAC	CAC	AAC	AAT	AAC	CAG	TTT	TTC	CAC	GAT	GTG	GAG	GTG	GAA	GAC	GTC	671	
Tyr	His	Asn	Asn	Asn	Gln	Phe	Phe	His	Asp	Val	Glu	Val	Glu	Asp	Val		
	1050				1055					1060					1065		
CTC	CCT	AGC	TCC	AAA	CGG	TCC	CCC	TCC	ACG	GCC	ACG	CTG	AGT	CTG	CCC	719	
Leu	Pro	Ser	Ser	Lys	Arg	Ser	Pro	Ser	Thr	Ala	Thr	Leu	Ser	Leu	Pro		
				1070					1075					1080			
AGC	CTG	GAG	GCC	TAC	AGA	GAC	CCC	TCG	TGC	CTG	AGC	CCG	GCC	AGC	AGC	767	
Ser	Leu	Glu	Ala	Tyr	Arg	Asp	Pro	Ser	Cys	Leu	Ser	Pro	Ala	Ser	Ser		
			1085					1090					1095				
CTG	TCC	TCC	CGG	AGC	TGC	AAC	TCA	GAG	GCC	TCC	TCC	TAC	GAG	TCC	AAC	815	
Leu	Ser	Ser	Arg	Ser	Cys	Asn	Ser	Glu	Ala	Ser	Ser	Tyr	Glu	Ser	Asn		
		1100					1105					1110					
TAC	TCG	TAC	CCG	TAC	GCG	TCC	CCC	CAG	ACG	TCG	CCA	TGG	CAG	TCT	CCC	863	
Tyr	Ser	Tyr	Pro	Tyr	Ala	Ser	Pro	Gln	Thr	Ser	Pro	Trp	Gln	Ser	Pro		
	1115					1120					1125						
TGC	GTG	TCT	CCC	AAG	ACC	ACG	GAC	CCC	GAG	GAG	GGC	TTT	CCC	CGC	GGG	911	
Cys	Val	Ser	Pro	Lys	Thr	Thr	Asp	Pro	Glu	Glu	Gly	Phe	Pro	Arg	Gly		
	1130				1135					1140					1145		
CTG	GGG	GCC	TGC	ACA	CTG	CTG	GGT	TCC	CCG	CAG	CAC	TCC	CCC	TCC	ACC	959	
Leu	Gly	Ala	Cys	Thr	Leu	Leu	Gly	Ser	Pro	Gln	His	Ser	Pro	Ser	Thr		
				1150					1155					1160			
TCG	CCC	CGC	GCC	AGC	GTC	ACT	GAG	GAG	AGC	TGG	CTG	GGT	GCC	CGC	TCC	1007	
Ser	Pro	Arg	Ala	Ser	Val	Thr	Glu	Glu	Ser	Trp	Leu	Gly	Ala	Arg	Ser		
			1165					1170					1175				
TCC	AGA	CCC	GCG	TCC	CCT	TGC	AAC	AAG	AGG	AAG	TAC	AGC	CTC	AAC	GGC	1055	
Ser	Arg	Pro	Ala	Ser	Pro	Cys	Asn	Lys	Arg	Lys	Tyr	Ser	Leu	Asn	Gly		
		1180					1185					1190					
CGG	CAG	CCG	CCC	TAC	TCA	CCC	CAC	CAC	TCG	CCC	ACG	CCG	TCC	CCG	CAC	1103	
Arg	Gln	Pro	Pro	Tyr	Ser	Pro	His	His	Ser	Pro	Thr	Pro	Ser	Pro	His		
	1195					1200					1205						
GGC	TCC	CCG	CGG	GTC	AGC	GTG	ACC	GAC	GAC	TCG	TGG	TTG	GGC	AAC	ACC	1151	
Gly	Ser	Pro	Arg	Val	Ser	Val	Thr	Asp	Asp	Ser	Trp	Leu	Gly	Asn	Thr		
	1210				1215					1220					1225		
ACC	CAG	TAC	ACC	AGC	TCG	GCC	ATC	GTG	GCC	GCC	ATC	AAC	GCG	CTG	ACC	1199	
Thr	Gln	Tyr	Thr	Ser	Ser	Ala	Ile	Val	Ala	Ala	Ile	Asn	Ala	Leu	Thr		
				1230					1235					1240			
ACC	GAC	AGC	AGC	CTG	GAC	CTG	GGA	GAT	GGC	GTC	CCT	GTC	AAG	TCC	CGC	1247	
Thr	Asp	Ser	Ser	Leu	Asp	Leu	Gly	Asp	Gly	Val	Pro	Val	Lys	Ser	Arg		
			1245					1250					1255				
AAG	ACC	ACC	CTG	GAG	CAG	CCG	CCC	TCA	GTG	GCG	CTC	AAG	GTG	GAG	CCC	1295	
Lys	Thr	Thr	Leu	Glu	Gln	Pro	Pro	Ser	Val	Ala	Leu	Lys	Val	Glu	Pro		
		1260					1265					1270					
GTC	GGG	GAG	GAC	CTG	GGC	AGC	CCC	CCG	CCC	CCG	GCC	GAC	TTC	GCG	CCC	1343	
Val	Gly	Glu	Asp	Leu	Gly	Ser	Pro	Pro	Pro	Pro	Ala	Asp	Phe	Ala	Pro		
	1275					1280					1285						
GAA	GAC	TAC	TCC	TCT	TTC	CAG	CAC	ATC	AGG	AAG	GGC	GGC	TTC	TGC	GAC	1391	
Glu	Asp	Tyr	Ser	Ser	Phe	Gln	His	Ile	Arg	Lys	Gly	Gly	Phe	Cys	Asp		

1290	1295	1300	1305	
CAG TAC CTG GCG GTG CCG CAG CAC CCC TAC CAG TGG GCG AAG CCC AAG Gln Tyr Leu Ala Val Pro Gln His Pro Tyr Gln Trp Ala Lys Pro Lys 1310 1315 1320				1439
CCC CTG TCC CCT ACG TCC TAC ATG AGC CCG ACC CTG CCC GCC CTG GAC Pro Leu Ser Pro Thr Ser Tyr Met Ser Pro Thr Leu Pro Ala Leu Asp 1325 1330 1335				1487
TGG CAG CTG CCG TCC CAC TCA GGC CCG TAT GAG CTT CGG ATT GAG GTG Trp Gln Leu Pro Ser His Ser Gly Pro Tyr Glu Leu Arg Ile Glu Val 1340 1345 1350				1535
CAG CCC AAG TCC CAC CAC CGA GCC CAC TAC GAG ACG GAG GGC AGC CGG Gln Pro Lys Ser His His Arg Ala His Tyr Glu Thr Glu Gly Ser Arg 1355 1360 1365				1583
GGG GCC GTG AAG GCG TCG GCC GGA GGA CAC CCC ATC GTG CAG CTG CAT Gly Ala Val Lys Ala Ser Ala Gly Gly His Pro Ile Val Gln Leu His 1370 1375 1380 1385				1631
GGC TAC TTG GAG AAT GAG CCG CTG ATG CTG CAG CTT TTC ATT GGG ACG Gly Tyr Leu Glu Asn Glu Pro Leu Met Leu Gln Leu Phe Ile Gly Thr 1390 1395 1400				1679
GCG GAC GAC CGC CTG CTG CGC CCG CAC GCC TTC TAC CAG GTG CAC CGC Ala Asp Asp Arg Leu Leu Arg Pro His Ala Phe Tyr Gln Val His Arg 1405 1410 1415				1727
ATC ACA GGG AAG ACC GTG TCC ACC ACC AGC CAC GAG GCT ATC CTC TCC Ile Thr Gly Lys Thr Val Ser Thr Thr Ser His Glu Ala Ile Leu Ser 1420 1425 1430				1775
AAC ACC AAA GTC CTG GAG ATC CCA CTC CTG CCG GAG AAC AGC ATG CGA Asn Thr Lys Val Leu Glu Ile Pro Leu Leu Pro Glu Asn Ser Met Arg 1435 1440 1445				1823
GCC GTC ATT GAC TGT GCC GGA ATC CTG AAA CTC AGA AAC TCC GAC ATT Ala Val Ile Asp Cys Gly Ile Leu Lys Leu Arg Asn Ser Asp Ile 1450 1455 1460 1465				1871
GAA CTT CGG AAA GGA GAG ACG GAC ATC GGG AGG AAG AAC ACA CGG GTA Glu Leu Arg Lys Gly Glu Thr Asp Ile Gly Arg Lys Asn Thr Arg Val 1470 1475 1480				1919
CGG CTG GTG TTC CGC GTT CAC GTC CCG CAA CCC AGC GGC CGC ACG CTG Arg Leu Val Phe Arg Val His Val Pro Gln Pro Ser Gly Arg Thr Leu 1485 1490 1495				1967
TCC CTG CAG GTG GCC TCC AAC CCC ATC GAA TGC TCC CAG CGC TCA GCT Ser Leu Gln Val Ala Ser Asn Pro Ile Glu Cys Ser Gln Arg Ser Ala 1500 1505 1510				2015
CAG GAG CTG CCT CTG GTG GAG AAG CAG AGC ACG GAC AGC TAT CCG GTC Gln Glu Leu Pro Leu Val Glu Lys Gln Ser Thr Asp Ser Tyr Pro Val 1515 1520 1525				2063
GTG GGC GGG AAG AAG ATG GTC CTG TCT GGC CAC AAC TTC CTG CAG GAC Val Gly Gly Lys Lys Met Val Leu Ser Gly His Asn Phe Leu Gln Asp 1530 1535 1540 1545				2111
TCC AAG GTC ATT TTC GTG GAG AAA GCC CCA GAT GGC CAC CAT GTC TGG Ser Lys Val Ile Phe Val Glu Lys Ala Pro Asp Gly His His Val Trp 1550 1555 1560				2159
GAG ATG GAA GCG AAA ACT GAC CGG GAC CTG TGC AAG CCG AAT TCT CTG Glu Met Glu Ala Lys Thr Asp Arg Asp Leu Cys Lys Pro Asn Ser Leu 1565 1570 1575				2207

GTG GTT GAG ATC CCG CCA TTT CGG AAT CAG AGG ATA ACC AGC CCC GTT	2255
Val Val Glu Ile Pro Pro Phe Arg Asn Gln Arg Ile Thr Ser Pro Val	
1580 1585 1590	
CAC GTC AGT TTC TAC GTC TGC AAC GGG AAG AGA AAG CGA AGC CAG TAC	2303
His Val Ser Phe Tyr Val Cys Asn Gly Lys Arg Lys Arg Ser Gln Tyr	
1595 1600 1605	
CAG CGT TTC ACC TAC CTT CCC GCC AAC GGT AAC GCC ATC TTT CTA ACC	2351
Gln Arg Phe Thr Tyr Leu Pro Ala Asn Gly Asn Ala Ile Phe Leu Thr	
1610 1615 1620 1625	
GTA AGC CGT GAA CAT GAG CGC GTG GGG TGC TTT TTC TAA AGACGCAGAA	2400
Val Ser Arg Glu His Glu Arg Val Gly Cys Phe Phe *	
1630 1635	
ACGACGTCGC CGTAAAGCAG CGTGGCGTGT TGCACATTTA ACTGTGTGAT GTCCCGTTAG	2460
TGAGACCGAG CCATCGATGC CCTGAAAAGG AAAGGAAAAG GGAAGCTTCG GATGCATTTT	2520
CCTTGATCCC TGTTGGGGGT GGGGGGCGGG GGTTCATAC TCAGATAGTC ACGGTTATTT	2580
TGCTTCTTGC GAATGTATAA CAGCCAAGGG GAAAACATGG CTCTTCTGCT CCAAAAAACT	2640
GAGGGGGTCC TGGTGTGCAT TTGCACCTTA AAGCTGCTTA CGGTGAAAAG GCAAATAGGT	2700
ATAGCTATTT TGCAGGCACC TTTAGGAATA AACTTTGCTT TTA	2743

(2) INFORMATION FOR SEQ ID NO:4:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 717 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

Met Pro Ser Thr Ser Phe Pro Val Pro Ser Lys Phe Pro Leu Gly Pro	
1 5 10 15	
Ala Ala Ala Val Phe Gly Arg Gly Glu Thr Leu Gly Pro Ala Pro Arg	
20 25 30	
Ala Gly Gly Thr Met Lys Ser Ala Glu Glu Glu His Tyr Gly Tyr Ala	
35 40 45	
Ser Ser Asn Val Ser Pro Ala Leu Pro Leu Pro Thr Ala His Ser Thr	
50 55 60	
Leu Pro Ala Pro Cys His Asn Leu Gln Thr Ser Thr Pro Gly Ile Ile	
65 70 75 80	
Pro Pro Ala Asp His Pro Ser Gly Tyr Gly Ala Ala Leu Asp Gly Gly	
85 90 95	
Pro Ala Gly Tyr Phe Leu Ser Ser Gly His Thr Arg Pro Asp Gly Ala	
100 105 110	
Pro Ala Leu Glu Ser Pro Arg Ile Glu Ile Thr Ser Cys Leu Gly Leu	
115 120 125	
Tyr His Asn Asn Asn Gln Phe Phe His Asp Val Glu Val Glu Asp Val	
130 135 140	
Leu Pro Ser Ser Lys Arg Ser Pro Ser Thr Ala Thr Leu Ser Leu Pro	
145 150 155 160	

Ser Leu Glu Ala Tyr Arg Asp Pro Ser Cys Leu Ser Pro Ala Ser Ser
 165 170 175
 Leu Ser Ser Arg Ser Cys Asn Ser Glu Ala Ser Ser Tyr Glu Ser Asn
 180 185 190
 Tyr Ser Tyr Pro Tyr Ala Ser Pro Gln Thr Ser Pro Trp Gln Ser Pro
 195 200 205
 Cys Val Ser Pro Lys Thr Thr Asp Pro Glu Glu Gly Phe Pro Arg Gly
 210 215 220
 Leu Gly Ala Cys Thr Leu Leu Gly Ser Pro Gln His Ser Pro Ser Thr
 225 230 235 240
 Ser Pro Arg Ala Ser Val Thr Glu Glu Ser Trp Leu Gly Ala Arg Ser
 245 250 255
 Ser Arg Pro Ala Ser Pro Cys Asn Lys Arg Lys Tyr Ser Leu Asn Gly
 260 265 270
 Arg Gln Pro Pro Tyr Ser Pro His His Ser Pro Thr Pro Ser Pro His
 275 280 285
 Gly Ser Pro Arg Val Ser Val Thr Asp Asp Ser Trp Leu Gly Asn Thr
 290 295 300
 Thr Gln Tyr Thr Ser Ser Ala Ile Val Ala Ala Ile Asn Ala Leu Thr
 305 310 315 320
 Thr Asp Ser Ser Leu Asp Leu Gly Asp Gly Val Pro Val Lys Ser Arg
 325 330 335
 Lys Thr Thr Leu Glu Gln Pro Pro Ser Val Ala Leu Lys Val Glu Pro
 340 345 350
 Val Gly Glu Asp Leu Gly Ser Pro Pro Pro Pro Ala Asp Phe Ala Pro
 355 360 365
 Glu Asp Tyr Ser Ser Phe Gln His Ile Arg Lys Gly Gly Phe Cys Asp
 370 375 380
 Gln Tyr Leu Ala Val Pro Gln His Pro Tyr Gln Trp Ala Lys Pro Lys
 385 390 395 400
 Pro Leu Ser Pro Thr Ser Tyr Met Ser Pro Thr Leu Pro Ala Leu Asp
 405 410 415
 Trp Gln Leu Pro Ser His Ser Gly Pro Tyr Glu Leu Arg Ile Glu Val
 420 425 430
 Gln Pro Lys Ser His His Arg Ala His Tyr Glu Thr Glu Gly Ser Arg
 435 440 445
 Gly Ala Val Lys Ala Ser Ala Gly Gly His Pro Ile Val Gln Leu His
 450 455 460
 Gly Tyr Leu Glu Asn Glu Pro Leu Met Leu Gln Leu Phe Ile Gly Thr
 465 470 475 480
 Ala Asp Asp Arg Leu Leu Arg Pro His Ala Phe Tyr Gln Val His Arg
 485 490 495
 Ile Thr Gly Lys Thr Val Ser Thr Thr Ser His Glu Ala Ile Leu Ser
 500 505 510
 Asn Thr Lys Val Leu Glu Ile Pro Leu Leu Pro Glu Asn Ser Met Arg
 515 520 525

Ala Val Ile Asp Cys Ala Gly Ile Leu Lys Leu Arg Asn Ser Asp Ile
530 535 540

Glu Leu Arg Lys Gly Glu Thr Asp Ile Gly Arg Lys Asn Thr Arg Val
545 550 555 560

Arg Leu Val Phe Arg Val His Val Pro Gln Pro Ser Gly Arg Thr Leu
565 570 575

Ser Leu Gln Val Ala Ser Asn Pro Ile Glu Cys Ser Gln Arg Ser Ala
580 585 590

Gln Glu Leu Pro Leu Val Glu Lys Gln Ser Thr Asp Ser Tyr Pro Val
595 600 605

Val Gly Gly Lys Lys Met Val Leu Ser Gly His Asn Phe Leu Gln Asp
610 615 620

Ser Lys Val Ile Phe Val Glu Lys Ala Pro Asp Gly His His Val Trp
625 630 635 640

Glu Met Glu Ala Lys Thr Asp Arg Asp Leu Cys Lys Pro Asn Ser Leu
645 650 655

Val Val Glu Ile Pro Pro Phe Arg Asn Gln Arg Ile Thr Ser Pro Val
660 665 670

His Val Ser Phe Tyr Val Cys Asn Gly Lys Arg Lys Arg Ser Gln Tyr
675 680 685

Gln Arg Phe Thr Tyr Leu Pro Ala Asn Gly Asn Ala Ile Phe Leu Thr
690 695 700

Val Ser Arg Glu His Glu Arg Val Gly Cys Phe Phe *

705 710 715

(2) INFORMATION FOR SEQ ID NO:5:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 2881 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: double
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 142..2850

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

GCTTCTGGAG GGAGGCGGCA GCGACGGAGG AGGGGGCTTC TCAGAGAAAG GGAGGGAGGG 60

AGCCACCCGG GTGAAGATAC AGCAGCCTCC TGAAGTCCCC CCTCCCACCC AGGCCGGGAC 120

CTGGGGGCTC CTGCCGGATC C ATG GGG GCG GCC AGC TGC GAG GAT GAG GAG 171

Met Gly Ala Ala Ser Cys Glu Asp Glu Glu 720 725

CTG GAA TTT AAG CTG GTG TTC GGG GAG GAA AAG GAG GCC CCC CCG CTG 219

Leu Glu Phe Lys Leu Val Phe Gly Glu Glu Lys Glu Ala Pro Pro Leu 730 735 740

GGC GCG GGG GGA TTG GGG GAA GAA CTG GAC TCA GAG GAT GCC CCG CCA 267

Gly Ala Gly Gly Leu Gly Glu Glu Leu Asp Ser Glu Asp Ala Pro Pro 745 750 755

TGC Cys 760	TGC Cys	CGT Arg	CTG Leu	GCC Ala	TTG Leu 765	GGA Gly	GAG Glu	CCC Pro	CCT Pro	CCC Pro 770	TAT Tyr	GGC Gly	GCT Ala	GCA Ala	CCT Pro 775	315
ATC Ile	GGT Gly	ATT Ile	CCC Pro	CGA Arg 780	CCT Pro	CCA Pro	CCC Pro	CCT Pro	CGG Arg 785	CCT Pro	GGC Gly	ATG Met	CAT His	TCG Ser 790	CCA Pro	363
CCG Pro	CCG Pro	CGA Arg	CCA Pro 795	GCC Ala	CCC Pro	TCA Ser	CCT Pro	GGC Gly 800	ACC Thr	TGG Trp	GAG Glu	AGC Ser	CAG Gln 805	CCC Pro	GCC Ala	411
AGG Arg	TCG Ser	GTG Val 810	AGG Arg	CTG Leu	GGA Gly	GGA Gly	CCA Pro 815	GGA Gly	GGG Gly	GGT Gly	GCT Ala	GGG Gly 820	GGT Gly	GCT Ala	GGG Gly	459
GGT Gly 825	GGC Gly	CGT Arg	GTT Val	CTC Leu	GAG Glu	TGT Cys 830	CCC Pro	AGC Ser	ATC Ile	CGC Arg	ATC Ile 835	ACC Thr	TCC Ser	ATC Ile	TCT Ser	507
CCC Pro 840	ACG Thr	CCG Pro	GAG Glu	CCG Pro	CCA Pro 845	GCA Ala	GCG Ala	CTG Leu	GAG Glu	GAC Asp 850	AAC Asn	CCT Pro	GAT Asp	GCC Ala	TGG Trp 855	555
GGG Gly	GAC Asp	GGC Gly	TCT Ser	CCT Pro 860	AGA Arg	GAT Asp	TAC Tyr	CCC Pro	CCA Pro 865	CCA Pro	GAA Glu	GGC Gly	TTT Phe	GGG Gly 870	GGC Gly	603
TAC Tyr	AGA Arg	GAA Glu	GCA Ala 875	GGG Gly	GCC Ala	CAG Gln	GGT Gly	GGG Gly 880	GGG Gly	GCC Ala	TTC Phe	TTC Phe	AGC Ser 885	CCA Pro	AGC Ser	651
CCT Pro	GGC Gly	AGC Ser 890	AGC Ser	AGC Ser	CTG Leu	TCC Ser	TCG Ser 895	TGG Trp	AGC Ser	TTC Phe	TTC Phe	TCC Ser 900	GAT Asp	GCC Ala	TCT Ser	699
GAC Asp 905	GAG Glu	GCA Ala	GCC Ala	CTG Leu	TAT Tyr	GCA Ala 910	GCC Ala	TGC Cys	GAC Asp	GAG Glu	GTG Val 915	GAG Glu	TCT Ser	GAG Glu	CTA Leu	747
AAT Asn 920	GAG Glu	GCG Ala	GCC Ala	TCC Ser	CGC Arg 925	TTT Phe	GGC Gly	CTG Leu	GGC Gly	TCC Ser 930	CCG Pro	CTG Leu	CCC Pro	TCG Ser	CCC Pro 935	795
CGG Arg	GCC Ala	TCC Ser	CCT Pro	CGG Arg 940	CCA Pro	TGG Trp	ACC Thr	CCC Pro	GAA Glu 945	GAT Asp	CCC Pro	TGG Trp	AGC Ser	CTG Leu 950	TAT Tyr	843
GGT Gly	CCA Pro	AGC Ser	CCC Pro 955	GGA Gly	GGC Gly	CGA Arg	GGG Gly	CCA Pro 960	GAG Glu	GAT Asp	AGC Ser	TGG Trp	CTA Leu 965	CTC Leu	CTC Leu	891
AGT Ser	GCT Ala	CCT Pro 970	GGG Gly	CCC Pro	ACC Thr	CCA Pro	GCC Ala 975	TCC Ser	CCG Pro	CGG Arg	CCT Pro	GCC Ala 980	TCT Ser	CCA Pro	TGT Cys	939
GGC Gly 985	AAG Lys	CGG Arg	CGC Arg	TAT Tyr	TCC Ser	AGC Ser 990	TCG Ser	GGA Gly	ACC Thr	CCA Pro	TCT Ser 995	TCA Ser	GCC Ala	TCC Ser	CCA Pro	987
GCT Ala 1000	CTG Leu	TCC Ser	CGC Arg	CGT Arg	GGC Gly 1005	AGC Ser	CTG Leu	GGG Gly	GAA Glu 1010	GAG Glu	GGG Gly	TCT Ser	GAG Glu	CCA Pro	CCT Pro 1015	1035
CCA Pro	CCA Pro	CCC Pro	CCA Pro	TTG Leu 1020	CCT Pro	CTG Leu	GCC Ala	CGG Arg	GAC Asp 1025	CCG Pro	GGC Gly	TCC Ser	CCT Pro	GGT Gly 1030	CCC Pro	1083
TTT Gly	GAC Ala	TAT Tyr	GTG Val	GGG Gly	GCC Ala	CCA Pro	CCA Pro	GCT Ala	GAG Glu	AGC Ser	ATC Ile	CCT Pro	CAG Gln	AAG Lys	ACA Thr	1131

45

1305	1310	1315	
GAA CTG GTA CTG ACC GGC TCC AAC TTC CTG CCA GAC TCC AAG GTG GTG Glu Leu Val Leu Thr Gly Ser Asn Phe Leu Pro Asp Ser Lys Val Val 1320 1325 1330 1335			1995
TTC ATT GAG AGG GGT CCT GAT GGG AAG CTG CAA TGG GAG GAG GAG GCC Phe Ile Glu Arg Gly Pro Asp Gly Lys Leu Gln Trp Glu Glu Glu Ala 1340 1345 1350			2043
ACA GTG AAC CGA CTG CAG AGC AAC GAG GTG ACG CTG ACC CTG ACT GTC Thr Val Asn Arg Leu Gln Ser Asn Glu Val Thr Leu Thr Leu Thr Val 1355 1360 1365			2091
CCC GAG TAC AGC AAC AAG AGG GTT TCC CGG CCA GTC CAG GTC TAC TTT Pro Glu Tyr Ser Asn Lys Arg Val Ser Arg Pro Val Gln Val Tyr Phe 1370 1375 1380			2139
TAT GTC TCC AAT GGG CGG AGG AAA CGC AGT CCT ACC CAG AGT TTC AGG Tyr Val Ser Asn Gly Arg Arg Lys Arg Ser Pro Thr Gln Ser Phe Arg 1385 1390 1395			2187
TTT CTG CCT GTG ATC TGC AAA GAG GAG CCC CTA CCG GAC TCA TCT CTG Phe Leu Pro Val Ile Cys Lys Glu Glu Pro Leu Pro Asp Ser Ser Leu 1400 1405 1410 1415			2235
CGG GGT TTC CCT TCA GCA TCG GCA ACC CCC TTT GGC ACT GAC ATG GAC Arg Gly Phe Pro Ser Ala Ser Ala Thr Pro Phe Gly Thr Asp Met Asp 1420 1425 1430			2283
TTC TCA CCA CCC AGG CCC CCC TAC CCC TCC TAT CCC CAT GAA GAC CCT Phe Ser Pro Pro Arg Pro Pro Tyr Pro Ser Tyr Pro His Glu Asp Pro 1435 1440 1445			2331
GCT TGC GAA ACT CCT TAC CTA TCA GAA GGC TTC GGC TAT GGC ATG CCC Ala Cys Glu Thr Pro Tyr Leu Ser Glu Gly Phe Gly Tyr Gly Met Pro 1450 1455 1460			2379
CCT CTG TAC CCC CAG ACG GGG CCC CCA CCA TCC TAC AGA CCG GGC CTG Pro Leu Tyr Pro Gln Thr Gly Pro Pro Pro Ser Tyr Arg Pro Gly Leu 1465 1470 1475			2427
CGG ATG TTC CCT GAG ACT AGG GGT ACC ACA GGT TGT GCC CAA CCA CCT Arg Met Phe Pro Glu Thr Arg Gly Thr Thr Gly Cys Ala Gln Pro Pro 1480 1485 1490 1495			2475
GCA GTT TCC TTC CTT CCC CGC CCC TTC CCT AGT GAC CCG TAT GGA GGG Ala Val Ser Phe Leu Pro Arg Pro Phe Pro Ser Asp Pro Tyr Gly Gly 1500 1505 1510			2523
CGG GGC TCC TCT TTC CCC CTG GGG CTG CCA TTC TCT CCG CCA GCC CCC Arg Gly Ser Ser Phe Pro Leu Gly Leu Pro Phe Ser Pro Pro Ala Pro 1515 1520 1525			2571
TTT CGG CCG CCT CCT CTT CCT GCA TCC CCA CCG CTT GAA GGC CCC TTC Phe Arg Pro Pro Pro Leu Pro Ala Ser Pro Pro Leu Glu Gly Pro Phe 1530 1535 1540			2619
CCT TCC CAG AGT GAT GTG CAT CCC CTA CCT GCT GAG GGA TAC AAT AAG Pro Ser Gln Ser Asp Val His Pro Leu Pro Ala Glu Gly Tyr Asn Lys 1545 1550 1555			2667
GTA GGG CCA GGC TAT GGC CCT GGG GAG GGG GCT CCG GAG CAG GAG AAA Val Gly Pro Gly Tyr Gly Pro Gly Glu Gly Ala Pro Glu Gln Glu Lys 1560 1565 1570 1575			2715
TCC AGG GGT GGC TAC AGC AGC GGC TTT CGA GAC AGT GTC CCT ATC CAG Ser Arg Gly Gly Tyr Ser Ser Gly Phe Arg Asp Ser Val Pro Ile Gln 1580 1585 1590			2763

GGT ATC ACG CTG GAG GAA GTG AGT GAG ATC ATT GGC CGA GAC CTG AGT 2811
 Gly Ile Thr Leu Glu Glu Val Ser Glu Ile Ile Gly Arg Asp Leu Ser
 1595 1600 1605

GGC TTC CCT GCA CCT CCT GGA GAA GAG CCT CCT GCC TGA ACCACGTGAA 2860
 Gly Phe Pro Ala Pro Pro Gly Glu Glu Pro Pro Ala *
 1610 1615 1620

CTGTCATCAC CTGGCAACCC C 2881

(2) INFORMATION FOR SEQ ID NO:6:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 903 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

Met Gly Ala Ala Ser Cys Glu Asp Glu Glu Leu Glu Phe Lys Leu Val
 1 5 10 15
 Phe Gly Glu Glu Lys Glu Ala Pro Pro Leu Gly Ala Gly Gly Leu Gly
 20 25 30
 Glu Glu Leu Asp Ser Glu Asp Ala Pro Pro Cys Cys Arg Leu Ala Leu
 35 40 45
 Gly Glu Pro Pro Pro Tyr Gly Ala Ala Pro Ile Gly Ile Pro Arg Pro
 50 55 60
 Pro Pro Pro Arg Pro Gly Met His Ser Pro Pro Pro Arg Pro Ala Pro
 65 70 75 80
 Ser Pro Gly Thr Trp Glu Ser Gln Pro Ala Arg Ser Val Arg Leu Gly
 85 90 95
 Gly Pro Gly Gly Gly Ala Gly Gly Ala Gly Gly Gly Arg Val Leu Glu
 100 105 110
 Cys Pro Ser Ile Arg Ile Thr Ser Ile Ser Pro Thr Pro Glu Pro Pro
 115 120 125
 Ala Ala Leu Glu Asp Asn Pro Asp Ala Trp Gly Asp Gly Ser Pro Arg
 130 135 140
 Asp Tyr Pro Pro Pro Glu Gly Phe Gly Gly Tyr Arg Glu Ala Gly Ala
 145 150 155 160
 Gln Gly Gly Gly Ala Phe Phe Ser Pro Ser Pro Gly Ser Ser Ser Leu
 165 170 175
 Ser Ser Trp Ser Phe Phe Ser Asp Ala Ser Asp Glu Ala Ala Leu Tyr
 180 185 190
 Ala Ala Cys Asp Glu Val Glu Ser Glu Leu Asn Glu Ala Ala Ser Arg
 195 200 205
 Phe Gly Leu Gly Ser Pro Leu Pro Ser Pro Arg Ala Ser Pro Arg Pro
 210 215 220
 Trp Thr Pro Glu Asp Pro Trp Ser Leu Tyr Gly Pro Ser Pro Gly Gly
 225 230 235 240
 Arg Gly Pro Glu Asp Ser Trp Leu Leu Leu Ser Ala Pro Gly Pro Thr
 245 250 255

Pro Ala Ser Pro Arg Pro Ala Ser Pro Cys Gly Lys Arg Arg Tyr Ser
 260 265 270
 Ser Ser Gly Thr Pro Ser Ser Ala Ser Pro Ala Leu Ser Arg Arg Gly
 275 280 285
 Ser Leu Gly Glu Glu Gly Ser Glu Pro Pro Pro Pro Pro Pro Leu Pro
 290 295 300
 Leu Ala Arg Asp Pro Gly Ser Pro Gly Pro Phe Asp Tyr Val Gly Ala
 305 310 315 320
 Pro Pro Ala Glu Ser Ile Pro Gln Lys Thr Arg Arg Thr Ser Ser Glu
 325 330 335
 Gln Ala Val Ala Leu Pro Arg Ser Glu Glu Pro Ala Ser Cys Asn Gly
 340 345 350
 Lys Leu Pro Leu Gly Ala Glu Glu Ser Val Ala Pro Pro Gly Gly Ser
 355 360 365
 Arg Lys Glu Val Ala Gly Met Asp Tyr Leu Ala Val Pro Ser Pro Leu
 370 375 380
 Ala Trp Ser Lys Ala Arg Ile Gly Gly His Ser Pro Ile Phe Arg Thr
 385 390 395 400
 Ser Ala Leu Pro Pro Leu Asp Trp Pro Leu Pro Ser Gln Tyr Glu Gln
 405 410 415
 Leu Glu Leu Arg Ile Glu Val Gln Pro Arg Ala His His Arg Ala His
 420 425 430
 Tyr Glu Thr Glu Gly Ser Arg Gly Ala Val Lys Ala Ala Pro Gly Gly
 435 440 445
 His Pro Val Val Lys Leu Leu Gly Tyr Ser Glu Lys Pro Leu Thr Leu
 450 455 460
 Gln Met Phe Ile Gly Thr Ala Asp Glu Arg Asn Leu Arg Pro His Ala
 465 470 475 480
 Phe Tyr Gln Val His Arg Ile Thr Gly Lys Met Val Ala Thr Ala Ser
 485 490 495
 Tyr Glu Ala Val Val Ser Gly Thr Lys Val Leu Glu Met Thr Leu Leu
 500 505 510
 Pro Glu Asn Asn Met Ala Ala Asn Ile Asp Cys Ala Gly Ile Leu Lys
 515 520 525
 Leu Arg Asn Ser Asp Ile Glu Leu Arg Lys Gly Glu Thr Asp Ile Gly
 530 535 540
 Arg Lys Asn Thr Arg Val Arg Leu Val Phe Arg Val His Val Pro Gln
 545 550 555 560
 Gly Gly Gly Lys Val Val Ser Val Gln Ala Ala Ser Val Pro Ile Glu
 565 570 575
 Cys Ser Gln Arg Ser Ala Gln Glu Leu Pro Gln Val Glu Ala Tyr Ser
 580 585 590
 Pro Ser Ala Cys Ser Val Arg Gly Gly Glu Glu Leu Val Leu Thr Gly
 595 600 605
 Ser Asn Phe Leu Pro Asp Ser Lys Val Val Phe Ile Glu Arg Gly Pro
 610 615 620

Asp Gly Lys Leu Gln Trp Glu Glu Glu Ala Thr Val Asn Arg Leu Gln
 625 630 635 640
 Ser Asn Glu Val Thr Leu Thr Leu Thr Val Pro Glu Tyr Ser Asn Lys
 645 650 655
 Arg Val Ser Arg Pro Val Gln Val Tyr Phe Tyr Val Ser Asn Gly Arg
 660 665 670
 Arg Lys Arg Ser Pro Thr Gln Ser Phe Arg Phe Leu Pro Val Ile Cys
 675 680 685
 Lys Glu Glu Pro Leu Pro Asp Ser Ser Leu Arg Gly Phe Pro Ser Ala
 690 695 700
 Ser Ala Thr Pro Phe Gly Thr Asp Met Asp Phe Ser Pro Pro Arg Pro
 705 710 715 720
 Pro Tyr Pro Ser Tyr Pro His Glu Asp Pro Ala Cys Glu Thr Pro Tyr
 725 730 735
 Leu Ser Glu Gly Phe Gly Tyr Gly Met Pro Pro Leu Tyr Pro Gln Thr
 740 745 750
 Gly Pro Pro Pro Ser Tyr Arg Pro Gly Leu Arg Met Phe Pro Glu Thr
 755 760 765
 Arg Gly Thr Thr Gly Cys Ala Gln Pro Pro Ala Val Ser Phe Leu Pro
 770 775 780
 Arg Pro Phe Pro Ser Asp Pro Tyr Gly Gly Arg Gly Ser Ser Phe Pro
 785 790 795 800
 Leu Gly Leu Pro Phe Ser Pro Pro Ala Pro Phe Arg Pro Pro Pro Leu
 805 810 815
 Pro Ala Ser Pro Pro Leu Glu Gly Pro Phe Pro Ser Gln Ser Asp Val
 820 825 830
 His Pro Leu Pro Ala Glu Gly Tyr Asn Lys Val Gly Pro Gly Tyr Gly
 835 840 845
 Pro Gly Glu Gly Ala Pro Glu Gln Glu Lys Ser Arg Gly Gly Tyr Ser
 850 855 860
 Ser Gly Phe Arg Asp Ser Val Pro Ile Gln Gly Ile Thr Leu Glu Glu
 865 870 875 880
 Val Ser Glu Ile Ile Gly Arg Asp Leu Ser Gly Phe Pro Ala Pro Pro
 885 890 895
 Gly Glu Glu Pro Pro Ala *
 900

(2) INFORMATION FOR SEQ ID NO:7:

- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 2406 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 211..2337

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

CGGCTGCGGT TCCTGGTGCT GCTCGGCGCG CGGCCAGCTT TCGGAACGGA ACGCTCGGCG	60
TCGCGGGCCC CGCCCGGAAA GTTTGCCGTG GAGTCGCGAC CTCTTGGCCC GCGCGGCCCC	120
GCATGAAGCG GCGTTGAGGA GCTGCTGCCG CCGCTTGCCG CTGCCGCCGC CGCCGCCTGA	180
GGAGGAGCTG CAGCACCCTG GGCCACGCCG ATG ACT ACT GCA AAC TGT GGC GCC	234
Met Thr Thr Ala Asn Cys Gly Ala	905 910
CAC GAC GAG CTC GAC TTC AAA CTC GTC TTT GGC GAG GAC GGG GCG CCG	282
His Asp Glu Leu Asp Phe Lys Leu Val Phe Gly Glu Asp Gly Ala Pro	915 920 925
GCG CCG CCG CCC CCG GGC TCG CGG CCT GCA GAT CTT GAG CCA GAT GAT	330
Ala Pro Pro Pro Pro Gly Ser Arg Pro Ala Asp Leu Glu Pro Asp Asp	930 935 940
TGT GCA TCC ATT TAC ATC TTT AAT GTA GAT CCA CCT CCA TCT ACT TTA	378
Cys Ala Ser Ile Tyr Ile Phe Asn Val Asp Pro Pro Pro Ser Thr Leu	945 950 955
ACC ACA CCA CTT TGC TTA CCA CAT CAT GGA TTA CCG TCT CAC TCT TCT	426
Thr Thr Pro Leu Cys Leu Pro His His Gly Leu Pro Ser His Ser Ser	960 965 970 975
GTT TTG TCA CCA TCG TTT CAG CTC CAA AGT CAC AAA AAC TAT GAA GGA	474
Val Leu Ser Pro Ser Phe Gln Leu Gln Ser His Lys Asn Tyr Glu Gly	980 985 990
ACT TGT GAG ATT CCT GAA TCT AAA TAT AGC CCA TTA GGT GGT CCC AAA	522
Thr Cys Glu Ile Pro Glu Ser Lys Tyr Ser Pro Leu Gly Gly Pro Lys	995 1000 1005
CCC TTT GAG TGC CCA AGT ATT CAA ATT ACA TCT ATC TCT CCT AAC TGT	570
Pro Phe Glu Cys Pro Ser Ile Gln Ile Thr Ser Ile Ser Pro Asn Cys	1010 1015 1020
CAT CAA GAA TTA GAT GCA CAT GAA GAT GAC CTA CAG ATA AAT GAC CCA	618
His Gln Glu Leu Asp Ala His Glu Asp Asp Leu Gln Ile Asn Asp Pro	1025 1030 1035
GAA CGG GAA TTT TTG GAA AGG CCT TCT AGA GAT CAT CTC TAT CTT CCT	666
Glu Arg Glu Phe Leu Glu Arg Pro Ser Arg Asp His Leu Tyr Leu Pro	1040 1045 1050 1055
CTT GAG CCA TCC TAC CGG GAG TCT TCT CTT AGT CCT AGT CCT GCC AGC	714
Leu Glu Pro Ser Tyr Arg Glu Ser Ser Leu Ser Pro Ser Pro Ala Ser	1060 1065 1070
AGC ATC TCT TCT AGG AGT TGG TTC TCT GAT GCA TCT TCT TGT GAA TCG	762
Ser Ile Ser Ser Arg Ser Trp Phe Ser Asp Ala Ser Ser Cys Glu Ser	1075 1080 1085
CTT TCA CAT ATT TAT GAT GAT GTG GAC TCA GAG TTG AAT GAA GCT GCA	810
Leu Ser His Ile Tyr Asp Asp Val Asp Ser Glu Leu Asn Glu Ala Ala	1090 1095 1100
GCC CGA TTT ACC CTT GGA TCC CCT CTG ACT TCT CCT GGT GGC TCT CCA	858
Ala Arg Phe Thr Leu Gly Ser Pro Leu Thr Ser Pro Gly Gly Ser Pro	1105 1110 1115
GGG GGC TGC CCT GGA GAA GAA ACT TGG CAT CAA CAG TAT GGA CTT GGA	906
Gly Gly Cys Pro Gly Glu Glu Thr Trp His Gln Gln Tyr Gly Leu Gly	1120 1125 1130 1135
CAC TCA TTA TCA CCC AGG CAA TCT CCT TGC CAC TCT CCT AGA TCC AGT	954

His	Ser	Leu	Ser	Pro	Arg	Gln	Ser	Pro	Cys	His	Ser	Pro	Arg	Ser	Ser		
				1140					1145						1150		
GTC	ACT	GAT	GAG	AAT	TGG	CTG	AGC	CCC	AGG	CCA	GCC	TCA	GGA	CCC	TCA	1002	
Val	Thr	Asp	Glu	Asn	Trp	Leu	Ser	Pro	Arg	Pro	Ala	Ser	Gly	Pro	Ser		
			1155					1160					1165				
TCA	AGG	CCC	ACA	TCC	CCC	TGT	GGG	AAA	CGG	AGG	CAC	TCC	AGT	GCT	GAA	1050	
Ser	Arg	Pro	Thr	Ser	Pro	Cys	Gly	Lys	Arg	Arg	His	Ser	Ser	Ala	Glu		
			1170				1175					1180					
GTT	TGT	TAT	GCT	GGG	TCC	CTT	TCA	CCC	CAT	CAC	TCA	CCT	GTT	CCT	TCA	1098	
Val	Cys	Tyr	Ala	Gly	Ser	Leu	Ser	Pro	His	His	Ser	Pro	Val	Pro	Ser		
	1185					1190						1195					
CCT	GGT	CAC	TCC	CCC	AGG	GGA	AGT	GTG	ACA	GAA	GAT	ACG	TGG	CTC	AAT	1146	
Pro	Gly	His	Ser	Pro	Arg	Gly	Ser	Val	Thr	Glu	Asp	Thr	Trp	Leu	Asn		
1200					1205					1210					1215		
GCT	TCT	GTC	CAT	GGT	GGG	TCA	GGC	CTT	GGC	CCT	GCA	GTT	TTT	CCA	TTT	1194	
Ala	Ser	Val	His	Gly	Gly	Ser	Gly	Leu	Gly	Pro	Ala	Val	Phe	Pro	Phe		
				1220				1225						1230			
CAG	TAC	TGT	GTA	GAG	ACT	GAC	ATC	CCT	CTC	AAA	ACA	AGG	AAA	ACT	TCT	1242	
Gln	Tyr	Cys	Val	Glu	Thr	Asp	Ile	Pro	Leu	Lys	Thr	Arg	Lys	Thr	Ser		
			1235					1240					1245				
GAA	GAT	CAA	GCT	GCC	ATA	CTA	CCA	GGA	AAA	TTA	GAG	CTG	TGT	TCA	GAT	1290	
Glu	Asp	Gln	Ala	Ala	Ile	Leu	Pro	Gly	Lys	Leu	Glu	Leu	Cys	Ser	Asp		
		1250					1255					1260					
GAC	CAA	GGG	AGT	TTA	TCA	CCA	GCC	CGG	GAG	ACT	TCA	ATA	GAT	GAT	GGC	1338	
Asp	Gln	Gly	Ser	Leu	Ser	Pro	Ala	Arg	Glu	Thr	Ser	Ile	Asp	Asp	Gly		
	1265					1270					1275						
CTT	GGA	TCT	CAG	TAT	CCT	TTA	AAG	AAA	GAT	TCA	TGT	GGT	GAT	CAG	TTT	1386	
Leu	Gly	Ser	Gln	Tyr	Pro	Leu	Lys	Lys	Asp	Ser	Cys	Gly	Asp	Gln	Phe		
1280					1285				1290					1295			
CTT	TCA	GTT	CCT	TCA	CCC	TTT	ACC	TGG	AGC	AAA	CCA	AAG	CCT	GGC	CAC	1434	
Leu	Ser	Val	Pro	Ser	Pro	Phe	Thr	Trp	Ser	Lys	Pro	Lys	Pro	Gly	His		
			1300					1305						1310			
ACC	CCT	ATA	TTT	CGC	ACA	TCT	TCA	TTA	CCT	CCA	CTA	GAC	TGG	CCT	TTA	1482	
Thr	Pro	Ile	Phe	Arg	Thr	Ser	Ser	Leu	Pro	Pro	Leu	Asp	Trp	Pro	Leu		
			1315					1320					1325				
CCA	GCT	CAT	TTT	GGA	CAA	TGT	GAA	CTG	AAA	ATA	GAA	GTG	CAA	CCT	AAA	1530	
Pro	Ala	His	Phe	Gly	Gln	Cys	Glu	Leu	Lys	Ile	Glu	Val	Gln	Pro	Lys		
		1330					1335					1340					
ACT	CAT	CAT	CGA	GCC	CAT	TAT	GAA	ACT	GAA	GGT	AGC	CGA	GGG	GCA	GTA	1578	
Thr	His	His	Arg	Ala	His	Tyr	Glu	Thr	Glu	Gly	Ser	Arg	Gly	Ala	Val		
	1345					1350					1355						
AAA	GCA	TCT	ACT	GGG	GGA	CAT	CCT	GTT	GTG	AAG	CTC	CTG	GGC	TAT	AAC	1626	
Lys	Ala	Ser	Thr	Gly	Gly	His	Pro	Val	Val	Lys	Leu	Leu	Gly	Tyr	Asn		
1360					1365					1370				1375			
GAA	AAG	CCA	ATA	AAT	CTA	CAA	ATG	TTT	ATT	GGG	ACA	GCA	GAT	GAT	CGA	1674	
Glu	Lys	Pro	Ile	Asn	Leu	Gln	Met	Phe	Ile	Gly	Thr	Ala	Asp	Asp	Arg		
			1380					1385						1390			
TAT	TTA	CGA	CCT	CAT	GCA	TTT	TAC	CAG	GTG	CAT	CGA	ATC	ACT	GGG	AAG	1722	
Tyr	Leu	Arg	Pro	His	Ala	Phe	Tyr	Gln	Val	His	Arg	Ile	Thr	Gly	Lys		
			1395					1400					1405				
ACA	GTC	GCT	ACT	GCA	AGC	CAA	GAG	ATA	ATA	ATT	GCC	AGT	ACA	AAA	GTT	1770	
Thr	Val	Ala	Thr	Ala	Ser	Gln	Glu	Ile	Ile	Ile	Ala	Ser	Thr	Lys	Val		

1410	1415	1420	
CTG GAA ATT CCA CTT CTT CCT GAA AAT AAT ATG TCA GCC AGT ATT GAT Leu Glu Ile Pro Leu Leu Pro Glu Asn Asn Met Ser Ala Ser Ile Asp 1425 1430 1435			1818
TGT GCA GGT ATT TTG AAA CTC CGC AAT TCA GAT ATA GAA CTT CGA AAA Cys Ala Gly Ile Leu Lys Leu Arg Asn Ser Asp Ile Glu Leu Arg Lys 1440 1445 1450 1455			1866
GGA GAA ACT GAT ATT GGC AGA AAG AAT ACT AGA GTA CGA CTT GTG TTT Gly Glu Thr Asp Ile Gly Arg Lys Asn Thr Arg Val Arg Leu Val Phe 1460 1465 1470			1914
CGT GTA CAC ATC CCA CAG CCC AGT GGA AAA GTC CTT TCT CTG CAG ATA Arg Val His Ile Pro Gln Pro Ser Gly Lys Val Leu Ser Leu Gln Ile 1475 1480 1485			1962
GCC TCT ATA CCC GTT GAG TGC TCC CAG CGG TCT GCT CAA GAA CTT CCT Ala Ser Ile Pro Val Glu Cys Ser Gln Arg Ser Ala Gln Glu Leu Pro 1490 1495 1500			2010
CAT ATT GAG AAG TAC AGT ATC AAC AGT TGT TCT GTA AAT GGA GGT CAT His Ile Glu Lys Tyr Ser Ile Asn Ser Cys Ser Val Asn Gly Gly His 1505 1510 1515			2058
GAA ATG GTT GTG ACT GGA TCT AAT TTT CTT CCA GAA TCC AAA ATC ATT Glu Met Val Val Thr Gly Ser Asn Phe Leu Pro Glu Ser Lys Ile Ile 1520 1525 1530 1535			2106
TTT CTT GAA AAA GGA CAA GAT GGA CGA CCT CAG TGG GAG GTA GAA GGG Phe Leu Glu Lys Gly Gln Asp Gly Arg Pro Gln Trp Glu Val Glu Gly 1540 1545 1550			2154
AAG ATA ATC AGG GAA AAA TGT CAA GGG GCT CAC ATT GTC CTT GAA GTT Lys Ile Ile Arg Glu Lys Cys Gln Gly Ala His Ile Val Leu Glu Val 1555 1560 1565			2202
CCT CCA TAT CAT AAC CCA GCA GTT ACA GCT GCA GTG CAG GTG CAC TTT Pro Pro Tyr His Asn Pro Ala Val Thr Ala Ala Val Gln Val His Phe 1570 1575 1580			2250
TAT CTT TGC AAT GGC AAG AGG AAA AAA AGC CAG TCT CAA CGT TTT ACT Tyr Leu Cys Asn Gly Lys Arg Lys Lys Ser Gln Ser Gln Arg Phe Thr 1585 1590 1595			2298
TAT ACA CCA GGT ACG AGG AGT CAT GAT GGT TTA CTA TAG AGCTTTCTTT Tyr Thr Pro Gly Thr Arg Ser His Asp Gly Leu Leu * 1600 1605 1610			2347
CCTAATGAAT AAAAAGTTAT TTAACGAACA AAAAAAAAAA AAAAAAAAAA AAAAAAAAAA			2406

(2) INFORMATION FOR SEQ ID NO:8:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 709 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

Met	Thr	Thr	Ala	Asn	Cys	Gly	Ala	His	Asp	Glu	Leu	Asp	Phe	Lys	Leu
1				5					10					15	
Val	Phe	Gly	Glu	Asp	Gly	Ala	Pro	Ala	Pro	Pro	Pro	Gly	Ser	Arg	
		20						25					30		

Pro Ala Asp Leu Glu Pro Asp Asp Cys Ala Ser Ile Tyr Ile Phe Asn
 35 40 45
 Val Asp Pro Pro Pro Ser Thr Leu Thr Thr Pro Leu Cys Leu Pro His
 50 55 60
 His Gly Leu Pro Ser His Ser Ser Val Leu Ser Pro Ser Phe Gln Leu
 65 70 75 80
 Gln Ser His Lys Asn Tyr Glu Gly Thr Cys Glu Ile Pro Glu Ser Lys
 85 90 95
 Tyr Ser Pro Leu Gly Gly Pro Lys Pro Phe Glu Cys Pro Ser Ile Gln
 100 105 110
 Ile Thr Ser Ile Ser Pro Asn Cys His Gln Glu Leu Asp Ala His Glu
 115 120 125
 Asp Asp Leu Gln Ile Asn Asp Pro Glu Arg Glu Phe Leu Glu Arg Pro
 130 135 140
 Ser Arg Asp His Leu Tyr Leu Pro Leu Glu Pro Ser Tyr Arg Glu Ser
 145 150 155 160
 Ser Leu Ser Pro Ser Pro Ala Ser Ser Ile Ser Ser Arg Ser Trp Phe
 165 170 175
 Ser Asp Ala Ser Ser Cys Glu Ser Leu Ser His Ile Tyr Asp Asp Val
 180 185 190
 Asp Ser Glu Leu Asn Glu Ala Ala Ala Arg Phe Thr Leu Gly Ser Pro
 195 200 205
 Leu Thr Ser Pro Gly Gly Ser Pro Gly Gly Cys Pro Gly Glu Glu Thr
 210 215 220
 Trp His Gln Gln Tyr Gly Leu Gly His Ser Leu Ser Pro Arg Gln Ser
 225 230 235 240
 Pro Cys His Ser Pro Arg Ser Ser Val Thr Asp Glu Asn Trp Leu Ser
 245 250 255
 Pro Arg Pro Ala Ser Gly Pro Ser Ser Arg Pro Thr Ser Pro Cys Gly
 260 265 270
 Lys Arg Arg His Ser Ser Ala Glu Val Cys Tyr Ala Gly Ser Leu Ser
 275 280 285
 Pro His His Ser Pro Val Pro Ser Pro Gly His Ser Pro Arg Gly Ser
 290 295 300
 Val Thr Glu Asp Thr Trp Leu Asn Ala Ser Val His Gly Gly Ser Gly
 305 310 315 320
 Leu Gly Pro Ala Val Phe Pro Phe Gln Tyr Cys Val Glu Thr Asp Ile
 325 330 335
 Pro Leu Lys Thr Arg Lys Thr Ser Glu Asp Gln Ala Ala Ile Leu Pro
 340 345 350
 Gly Lys Leu Glu Leu Cys Ser Asp Asp Gln Gly Ser Leu Ser Pro Ala
 355 360 365
 Arg Glu Thr Ser Ile Asp Asp Gly Leu Gly Ser Gln Tyr Pro Leu Lys
 370 375 380
 Lys Asp Ser Cys Gly Asp Gln Phe Leu Ser Val Pro Ser Pro Phe Thr
 385 390 395 400

Trp Ser Lys Pro Lys Pro Gly His Thr Pro Ile Phe Arg Thr Ser Ser
 405 410 415
 Leu Pro Pro Leu Asp Trp Pro Leu Pro Ala His Phe Gly Gln Cys Glu
 420 425 430
 Leu Lys Ile Glu Val Gln Pro Lys Thr His His Arg Ala His Tyr Glu
 435 440 445
 Thr Glu Gly Ser Arg Gly Ala Val Lys Ala Ser Thr Gly Gly His Pro
 450 455 460
 Val Val Lys Leu Leu Gly Tyr Asn Glu Lys Pro Ile Asn Leu Gln Met
 465 470 475 480
 Phe Ile Gly Thr Ala Asp Asp Arg Tyr Leu Arg Pro His Ala Phe Tyr
 485 490 495
 Gln Val His Arg Ile Thr Gly Lys Thr Val Ala Thr Ala Ser Gln Glu
 500 505 510
 Ile Ile Ile Ala Ser Thr Lys Val Leu Glu Ile Pro Leu Leu Pro Glu
 515 520 525
 Asn Asn Met Ser Ala Ser Ile Asp Cys Ala Gly Ile Leu Lys Leu Arg
 530 535 540
 Asn Ser Asp Ile Glu Leu Arg Lys Gly Glu Thr Asp Ile Gly Arg Lys
 545 550 555 560
 Asn Thr Arg Val Arg Leu Val Phe Arg Val His Ile Pro Gln Pro Ser
 565 570 575
 Gly Lys Val Leu Ser Leu Gln Ile Ala Ser Ile Pro Val Glu Cys Ser
 580 585 590
 Gln Arg Ser Ala Gln Glu Leu Pro His Ile Glu Lys Tyr Ser Ile Asn
 595 600 605
 Ser Cys Ser Val Asn Gly Gly His Glu Met Val Val Thr Gly Ser Asn
 610 615 620
 Phe Leu Pro Glu Ser Lys Ile Ile Phe Leu Glu Lys Gly Gln Asp Gly
 625 630 635 640
 Arg Pro Gln Trp Glu Val Glu Gly Lys Ile Ile Arg Glu Lys Cys Gln
 645 650 655
 Gly Ala His Ile Val Leu Glu Val Pro Pro Tyr His Asn Pro Ala Val
 660 665 670
 Thr Ala Ala Val Gln Val His Phe Tyr Leu Cys Asn Gly Lys Arg Lys
 675 680 685
 Lys Ser Gln Ser Gln Arg Phe Thr Tyr Thr Pro Gly Thr Arg Ser His
 690 695 700
 Asp Gly Leu Leu *
 705

(2) INFORMATION FOR SEQ ID NO:9:

- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 340 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

```

GTTTTGATGA AGCAAGAACA CAGAGAAGAG ATTGATTTGT CTTAGTTCC AACTTTGCCA      60
CAGACCTCTC GGCAAACCTCT GCTCGGGTCT CAGCCTCCTT CAGCTTCTCC TCCAACAGTT    120
TGATCTCCTC TTCATATTTA TCTTCTTTGG TGAATACTT GTCCGCCTGG GCCTCCAGGG      180
ATTTCAAGTT GTTGTAACA ATTTTCAGCT CCTCCTCTAG GTCCCCACAT TTAATCTCGG      240
CCACCTCAGC CCTCTCCTCC GAGCGCTCCA GCTCTCCTTC CAGGATCACC AGCTTCCTGG      300
CCACCTCTTC ATATTTGCGG TCTGAATCCT CAGCGATGTG      340

```

(2) INFORMATION FOR SEQ ID NO:10:

- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 40 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

```

Val Leu Met Lys Gln Glu His Arg Glu Glu Ile Asp Leu Ser Ser Val
1           5           10           15
Pro Thr Leu Pro Gln Thr Ser Arg Gln Thr Leu Leu Gly Ser Gln Pro
          20           25           30
Pro Ser Ala Ser Pro Pro Thr Val
          35           40

```

(2) INFORMATION FOR SEQ ID NO:11:

- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 1662 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

```

GTTTTGATGA AGCAAGAACA CAGAGAAGAG ATTGATTTGT CTTAGTTCC ATCTTTGCCT      60
GTGCCTCATC CTGCTCAGAC CCAGAGGCCT TCCTCTGATT CAGGCTGTTC ACATGACAGT    120
GTAATGTCAG GACAGAGAAG TTTGATTTGC TCCATCCAC AACATATGC ATCCATGGTG      180
ACCTCATCCC ATCTGCCACA GTTGCAAGTGT AGAGATGAGA GTGTTAGTAA AGAACAGCAT      240
ATGATTCCTT CTCCAATTGT ACACCAGCCT TTTCAAGTCA CACCAACACC TCCTGTGGGG      300
TCTTCCTATC AGCCTATGCA AACTAATGTT GTGTACAATG GACCAACTTG TCTTCCTATT      360
AATGCTGCCT CTAGTCAAGA ATTTGATTCA GTTTGTGTTT AGCAGGATGC AACTCTTTCT      420

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GGTTTAGTGA ATCTTGGCTG TCAACCACTG TCATCCATAC CATTTCATTC TTCAAATTCA      480
GGCTCAACAG GACATCTCTT AGCCCATACA CCTCATTCTG TGCATACCCT GCCTCATCTG      540
CAATCAATGG GATATCATTG TTCAAATACA GGACAAAGAT CTCTTTCTTC TCCAGTGGCT      600
GACCAGATTA CAGGTCAGCC TTCGTCTCAG TTACAACCTA TTACATATGG TCCTTCACAT      660
TCAGGGTCTG CTACAACAGC TTCCCCAGCA GCTTCTCATC CCTTGGCTAG TTCACCGCTT      720
TCTGGGCCAC CATCTCCTCA GCTTCAGCCT ATGCCTTACC AATCTCCTAG CTCAGGAACT      780
GCCTCATCAC CGTCTCCAGC CACCAGAATG CATTCTGGAC AGCACTCAAC TCAAGCACAA      840
AGTACGGGCC AGGGGGGTCT TTCTGCACCT TCATCCTTAA TATGTCACAG TTTGTGTGAT      900
CCAGCGTCAT TTCCACCTGA TGGGGCAACT GTGAGCATTAA AACCTGAACC AGAAGATCGA      960
GAGCCTAACT TTGCAACCAT TGGTCTGCAG GACATCACTT TAGATGATGA CCAATTTATA     1020
TCTGACTTGG AACACCAGCC ATCAGGTTCA GCAGAGAAAT GGCCTAACCA CAGTGTGCTC     1080
TCATGTCCAG CTCCTTTCTG GAGAATCTAG AGGTGAACGA GATAATTGGG AGAGACATGT     1140
CCCAGATTTT TGTTTCCCAA GGAGCAGGGG TGAGCAGGCA GGCTCCCCCT CCGAGTCCTG     1200
AGTCCCTGGA TTTAGGAAGA TCTGATGGGC TCTAACAGTG CTTACTGCAG CCTTGTGTCC     1260
ACCACCAACT TCTCAGCATG TTTCTCTCCT TGGACCTTGG GTTTCCAACT CTGCAGCCTT     1320
CAGGTCTGGG GCCAGGAGTG GGACCCACCA TTTGTGGGGA AAGTAGCATT CCTCCACCTC     1380
AGGCCTTGGG TAGATTTGGC AAAAGAACAG GAGCAGCATA GGCTGTTTGA GCTTTGGGGA     1440
AATGAACTTT GCTTTTTTATA TTAACTAGG ATACTTTTAT ATGATGGGTG CTTTGAGTGT     1500
GAATGCAGCA GGCTCTCTTG TTTCCGAGGT GCTGCTTTTG CAGGTGACCT GGTACTTATG     1560
CTAGGATTGG TGATTTGTAC TGCTTTATGG TCATTTGAAG GGCCCTTTAG TTTTATGAT     1620
AATTTTTTAAA ATAGGAACTT TTGATAAGAC CTTCTAGAAG CC                        1662

```

(2) INFORMATION FOR SEQ ID NO:12:

- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 369 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

```

Val Leu Met Lys Gln Glu His Arg Glu Glu Ile Asp Leu Ser Ser Val
1           5           10          15
Pro Ser Leu Pro Val Pro His Pro Ala Gln Thr Gln Arg Pro Ser Ser
20          25          30
Asp Ser Gly Cys Ser His Asp Ser Val Leu Ser Gly Gln Arg Ser Leu
35          40          45
Ile Cys Ser Ile Pro Gln Thr Tyr Ala Ser Met Val Thr Ser Ser His
50          55          60

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Leu Pro Gln Leu Gln Cys Arg Asp Glu Ser Val Ser Lys Glu Gln His
 65 70 75 80
 Met Ile Pro Ser Pro Ile Val His Gln Pro Phe Gln Val Thr Pro Thr
 85 90 95
 Pro Pro Val Gly Ser Ser Tyr Gln Pro Met Gln Thr Asn Val Val Tyr
 100 105 110
 Asn Gly Pro Thr Cys Leu Pro Ile Asn Ala Ala Ser Ser Gln Glu Phe
 115 120 125
 Asp Ser Val Leu Phe Gln Gln Asp Ala Thr Leu Ser Gly Leu Val Asn
 130 135 140
 Leu Gly Cys Gln Pro Leu Ser Ser Ile Pro Phe His Ser Ser Asn Ser
 145 150 155 160
 Gly Ser Thr Gly His Leu Leu Ala His Thr Pro His Ser Val His Thr
 165 170 175
 Leu Pro His Leu Gln Ser Met Gly Tyr His Cys Ser Asn Thr Gly Gln
 180 185 190
 Arg Ser Leu Ser Ser Pro Val Ala Asp Gln Ile Thr Gly Gln Pro Ser
 195 200 205
 Ser Gln Leu Gln Pro Ile Thr Tyr Gly Pro Ser His Ser Gly Ser Ala
 210 215 220
 Thr Thr Ala Ser Pro Ala Ala Ser His Pro Leu Ala Ser Ser Pro Leu
 225 230 235 240
 Ser Gly Pro Pro Ser Pro Gln Leu Gln Pro Met Pro Tyr Gln Ser Pro
 245 250 255
 Ser Ser Gly Thr Ala Ser Ser Pro Ser Pro Ala Thr Arg Met His Ser
 260 265 270
 Gly Gln His Ser Thr Gln Ala Gln Ser Thr Gly Gln Gly Gly Leu Ser
 275 280 285
 Ala Pro Ser Ser Leu Ile Cys His Ser Leu Cys Asp Pro Ala Ser Phe
 290 295 300
 Pro Pro Asp Gly Ala Thr Val Ser Ile Lys Pro Glu Pro Glu Asp Arg
 305 310 315 320
 Glu Pro Asn Phe Ala Thr Ile Gly Leu Gln Asp Ile Thr Leu Asp Asp
 325 330 335
 Asp Gln Phe Ile Ser Asp Leu Glu His Gln Pro Ser Gly Ser Ala Glu
 340 345 350
 Lys Trp Pro Asn His Ser Val Leu Ser Cys Pro Ala Pro Phe Trp Arg
 355 360 365
 Ile

(2) INFORMATION FOR SEQ ID NO:13:

- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 27 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

Asp Ile Glu Leu Arg Lys Gly Glu Thr Asp Ile Gly Arg Lys Asn Thr
1 5 10 15
Arg Val Arg Leu Val Phe Arg Val His Xaa Pro
20 25

(2) INFORMATION FOR SEQ ID NO:14:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 13 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:

Pro Xaa Glu Cys Ser Gln Arg Ser Ala Xaa Glu Leu Pro
1 5 10

(2) INFORMATION FOR SEQ ID NO:15:

(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 10 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:

GGAAAATTTT

10

(2) INFORMATION FOR SEQ ID NO:16:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 10 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:

GGAAAACTG

10

(2) INFORMATION FOR SEQ ID NO:17:

(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 23 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:

TACATTGGAA AATTTTATTA CAC

23

(2) INFORMATION FOR SEQ ID NO:18:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 30 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:18:

GGAGGAAAAA CTGTTTCATA CAGAAGGCGT

30

WHAT IS CLAIMED IS:

1. A human nuclear factor of activated T-cells, hNFAT, or fragment thereof having an hNFAT specific binding affinity.
- 5 2. A human nuclear factor of activated T-cells or fragment thereof according to claim 1, wherein said hNFAT is hNFATp₁ (SEQ ID NO:2).
3. A human nuclear factor of activated T-cells or fragment thereof according to
10 claim 1, wherein said hNFAT is hNFATp₂ (SEQ ID NO:2, residues 220-1021).
4. A human nuclear factor of activated T-cells or fragment thereof according to claim 1, wherein said hNFAT is hNFATc (SEQ ID NO:4).
- 15 5. A human nuclear factor of activated T-cells or fragment thereof according to claim 1, wherein said hNFAT is hNFAT3 (SEQ ID NO:6).
6. A human nuclear factor of activated T-cells or fragment thereof according to claim 1, wherein said hNFAT is hNFAT4a (SEQ ID NO:8).
- 20 7. A human nuclear factor of activated T-cells or fragment thereof according to claim 1, wherein said hNFAT is hNFAT4b (SEQ ID NO:8, residues 1-699 and SEQ ID NO:10).
- 25 8. A human nuclear factor of activated T-cells or fragment thereof according to claim 1, wherein said hNFAT is hNFAT4c (SEQ ID NO:8, residues 1-699 and SEQ ID NO:12).
9. A nucleic acid encoding a human nuclear factor of activated T-cells or fragment
30 thereof according to claim 1.

10. A method of identifying a pharmacological agent useful in the diagnosis or treatment of disease associated with the expression of a gene, wherein the expression of said gene is modulated by a transcription complex comprising a human nuclear factor of activated T-cells (hNFAT), said method comprising the steps of:
- 5 forming a mixture comprising a hNFAT or fragment thereof according to claim 1, a nucleic acid capable of selectively binding said hNFAT, a candidate pharmacological agent, and, optionally, a transcription factor different from said hNFAT or fragment thereof,;
- 10 incubating said mixture under conditions whereby, but for the presence of said candidate pharmacological agent, said hNFAT or fragment thereof selectively binds said nucleic acid and/or said hNFAT or fragment thereof, said transcription factor and said nucleic acid form a selectively bound complex;;
- 15 detecting the presence or absence of selective binding of said hNFAT or fragment thereof and said nucleic acid and/or said selectively bound complex;;
- wherein the absence of said selective binding and said selectively bound complex indicates that said candidate pharmacological agent is lead compound for a pharmacological agent capable of disrupting hNFAT dependent gene expression.

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US96/03113

A. CLASSIFICATION OF SUBJECT MATTER

IPC(6) : C07K 14/47; C12N 15/12; C12Q 1/68
US CL : 530/350; 536/23.5; 435/6

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 530/350; 536/23.5; 435/6

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

APS, DIALOG

search terms: NFAT, human, NFATp, NFATc, NFAT3, NFAT4, assay, transcription factor, binding, agent, compound, drug

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	NORTHROP et al. NF-AT components define a family of transcription factors targeted in T-cell activation. Nature. 09 June 1994, Vol. 369, pages 497-502, especially page 497.	1-4, 9
X	WO 94/15964 A1 (DANA-FARBER CANCER INSTITUTE, INC.) 21 July 1994, page 1, abstract; page 6, paragraph 2.	1-3, 9-10
X, P	WO 95/08554 A1 (THE BOARD OF TRUSTEES OF THE LELAND STANFORD JUNIOR UNIVERSITY) 30 March 1995, page 1, abstract; page 8, paragraph 2.	1, 4, 9-10
X	WO 95/02053 A1 (SCHERING CORPORATION) 19 January 1995, page 1, abstract; page 29, paragraph 2.	1-4, 6-10

☒ Further documents are listed in the continuation of Box C. ☐ See patent family annex.

* *A* *E* *L* *O* *P*	Special categories of cited documents: document defining the general state of the art which is not considered to be of particular relevance earlier document published on or after the international filing date document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified) document referring to an oral disclosure, use, exhibition or other means document published prior to the international filing date but later than the priority date claimed	*T* *X* *Y* *Z*	later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art document member of the same patent family
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Date of the actual completion of the international search

06 MAY 1996

Date of mailing of the international search report

03 JUN 1996

Name and mailing address of the ISA/US
Commissioner of Patents and Trademarks
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INTERNATIONAL SEARCH REPORT

International application No.
PCT/US96/03113

C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X, P	HOEY et al. Isolation of Two New Members of the NF-AT Gene Family and Functional Characterization of the NF-AT Proteins. Immunity. May 1995, Vol. 2, pages 461-472, especially page 461, abstract.	1, 5-9
A, P	HO et al. NFATc3, a Lymphoid-specific NFATc Family Member That Is Calcium-regulated and Exhibits Distinct DNA Binding Specificity. The Journal of Biological Chemistry. 25 August 1995, Vol. 270, No. 34, pages 19898-19907, see entire document.	1-10
A, P	MASUDA et al. NFATx, a Novel Member of the Nuclear Factor of Activated T Cells Family That Is Expressed Predominantly in the Thymus. May 1995, Vol. 15, No. 5, pages 2697-2706, see entire document.	1-10
A	MCCAFFREY et al. Isolation of the Cyclosporin-Sensitive T Cell Transcription Factor NFATp. Science. 29 October 1993, Vol. 262, pages 750-754, see entire document.	1-10

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